

23. Middlebrook, R., Hoegh-Guldberg, O. and Leggat, W., The effect of thermal history on the susceptibility of reef-building corals to thermal stress. *J. Exp. Biol.*, 2008, **211**, 1050–1056.
24. Hoegh-Guldberg, O. *et al.*, Coral reefs under rapid climate change and ocean acidification. *Science*, 2007, **318**, 1737–1742.
25. Chavanich, S., Viyakarn, V., Loyjiw, T., Pattaratamrong, P. and Chankong, A., Mass bleaching of soft coral, *Sarcophyton* spp. in Thailand and the role of temperature and salinity stress. *ICES J. Mar. Sci.*, 2009, **66**(7), 1515–1519.
26. Maynard, J. A., Anthony, K. R. N., Marshall, P. A. and Masiri, I., Major bleaching events can lead to increased thermal tolerance in corals. *Mar. Biol.*, 2008, **155**, 173–182.
27. Hoegh-Guldberg, O., Climate change and coral reefs: Trojan horse or false prophecy? *Coral Reefs*, 2008, **28**, 569–575.
28. Arthur, R., Coral Bleaching and mortality in three Indian reef regions during an El Niño southern oscillation event. *Curr. Sci.*, 2000, **79**, 1723–1729.

Received 30 June 2010; revised accepted 22 October 2010

Molecular taxonomy of marine mammals stranded along Kerala coast, India

Sanil George¹, K. Meenakshi¹ and A. Bijukumar^{2,*}

¹Chemical Biology Group, Rajiv Gandhi Centre for Biotechnology, Thiruvananthapuram 695 014, India

²Department of Aquatic Biology and Fisheries, University of Kerala, Thiruvananthapuram 695 581, India

Application of molecular tools for the identification of threatened marine mammals has gained importance in recent years. Though live and dead strandings of cetaceans are common along the Indian coasts, the specimens are often not properly identified due to the lack of local taxonomic expertise and poor quality of the specimens. Two marine mammals washed ashore in a putrefied condition at Edayar (08°25'N lat., 76°57'E long.), Thiruvananthapuram District, southwest coast of India, were identified by sequencing of 16S rRNA and COI genes. Sequence and phylogenetic similarity search done with all entries in the DNA sequence database, GenBank using BLAST identified the stranded mammals as Bryde's whale (*Balaenoptera edeni*) and finless porpoise (*Neophocaena phocaenoides*). The present report is the second record of *B. edeni* from the southwest coast of India.

Keywords: Cetaceans, molecular taxonomy, phylogeny, stranded mammals.

THE marine mammal diversity of Indian seas, represented by around 30 recorded species, forms almost one-fourth of the world's marine mammals, and almost 8% of all mammalian fauna recorded in India¹. The qualitative

deficiency of data on marine mammals in India notwithstanding, the shortcomings of marine mammal research in India include geographic disproportionateness in records, non-reporting of mortality due to fishing operations, lack of peer review, incorrect identification of species, incorrect geographic information, inaccuracy in measurements, repeated citation of incorrect records, misinterpretation of observations and lack of molecular data². Though live and dead strandings of marine mammals, especially cetaceans (whales, dolphins and porpoises), are common along the Indian coasts, the specimens are often not properly identified due to the lack of local taxonomic expertise and poor condition of the specimens. Since all the cetaceans are of importance from the conservation point of view, documenting their presence in the ecosystem and precise taxonomy would provide valuable information on various aspects of distribution and migratory nature of different species in the seas around India.

The application of molecular techniques has given stimulating impulses to marine mammal identification. In particular, partial or complete sequences of mitochondrial rRNA genes have been evaluated for appropriate evolutionary rates to resolve some aspects of the higher groups, such as genus, family and order. Because they contain information from old splitting events in their conserved regions, fast-evolving parts should be useful to resolve more recent events, e.g. intraspecific or intra-generic³. In addition, DNA sequences of mitochondrial cytochrome oxidase subunit I (COI) gene have been used to estimate phylogenetic relationships among closely related species^{4,5}. DNA sequencing technology has provided us the ability to determine the source of tissue samples believed to be derived from threatened or endangered species⁶. A phylogenetic approach to identifying marine mammal sequences from unknown sources has gained support in recent years⁷, especially for identifying whale meat products in the open market^{6,8}. Above all, sequencing of mitochondrial DNA has been used for describing new species of marine mammals^{9–11} and for describing their phylogeny¹². From India, initial efforts have been made by the Central Marine Fisheries Research Institute, Cochin towards molecular identification of marine mammals^{13–15}.

Two putrefied carcasses of marine mammals were washed ashore at Edayar (08°25'N lat., 76°57'E long.), Thiruvananthapuram District, southwest coast of India on 27 June 2009 (Figure 1 a and b). Based on field observations, though the presence of long ventral pleats on the lower profile of the head extending up to the navel confirmed the identity of one specimen as a whale belonging to the genus *Balaenoptera* (Bryde's whale, fin whale and blue whale in this group, recorded from India)¹⁶, confusion prevailed in the identification because of the dented upper jaw. Further, detailed examination of the specimen (total length = 3.9 m) also was difficult since the body was rotten, emanating foul odour, and the demand from

*For correspondence. (e-mail: abiju@rediffmail.com)



Figure 1. *Balaenoptera edeni* Anderson (a) and *Neophocaena phocaenoides* (Cuvier) (b) washed ashore at Edayar, Thiruvananthapuram.

Table 1. Primers used in the present study

Gene	Primer sequence	Annealing temperature (°C)	Product size (bp)	Source
16S	F – 5'-CGCCTGTTTATCAAAAACAT-3'	55	540	Palumbi <i>et al.</i> ¹⁷
rRNA	R – 5'-CCGGTCTGAACTCAGATCACGT-3'			
COI	F – 5'-GGTCAACAAATCATAAAGATATTG-3'	51	658	Folmer <i>et al.</i> ¹⁸
	R – 5'-TAAACTTCAGGGTGACCAAAAAATCA-3'			

the public (as the stranding was near a thickly inhabited area with many tourist resorts) was high to bury the carcass immediately. The second specimen (total length = 1.05 m) could not be identified precisely due to the collapsed head; blunt, laterally compressed teeth with expanded crowns and extremely short beak are the taxonomic characters distinguishing dolphins and porpoises¹⁶. Absence of pertinent taxonomic characters in the stranded marine mammals prompted us to opt for tools of molecular taxonomy.

The tissue samples collected from the marine mammals were processed for extraction of DNA using QIAGEN DNeasy Blood and Tissue kit, and amplified with 16S rRNA and COI genes in a 25 µl reaction volume with QIAGEN *Taq* PCR master mix kit using the thermal cycler Eppendorf. All the PCR products were visualized on 1% agarose gel and the most intense products were selected for sequencing. Primer details^{17,18} are given in Table 1. Sequencing was performed directly using the corresponding PCR primers and products were labelled using the BigDye Terminator V.3.1 Cycle sequencing kit (Applied Biosystems, Inc.) and sequenced using an ABI 3730 capillary sequencer following the manufacturer's instructions.

Sequence similarity search was done to identify the source species of the tissue, with all entries in the DNA sequence database GenBank, using Basic Local Alignment Search Tool (BLAST). In the case of the first specimen, BLAST search of COI showed 100% sequence

similarity with Bryde's whale (*Balaenoptera edeni* Anderson; family: Balaenopteridae), whereas in the case of the second specimen search of the 16S rRNA showed 97% sequence similarity with finless porpoise [*Neophocaena phocaenoides* (Cuvier); Family: Phocoenidae]. Though COI gene of *N. phocaenoides* was sequenced and the data were submitted to GenBank, further comparison was not possible due to absence of its COI sequence data in public domains and other publications.

Phylogenetic position of both the samples for 16S rRNA gene sequences was determined using neighbour-joining tree of Kimura-2-parameter distance model¹⁹, and the resultant clustering patterns are given in Figures 2–4. The results unequivocally showed that the first specimen belongs to *B. edeni* and the second to *N. phocaenoides*. The sequences were deposited in GenBank under accession numbers GQ856368 (16S rRNA of *B. edeni*), GQ856369 (16S rRNA of *N. phocaenoides*), GQ856370 (COI of *B. edeni*) and HQ268824 (COI of *N. phocaenoides*).

B. edeni occurring in the tropical, subtropical and warm temperate waters around the world is the least known of large baleen whales²⁰. The Bryde's whale is included in Appendix I of Conservation of International Trade of Endangered Species of Flora and Fauna (CITES) and listed in Appendix II of Convention on Migratory Species (CMS), and is thus offered protection at the international level. Among the whales, Bryde's whale is the least recorded from India, with only seven records from the country². The present report is the second from the

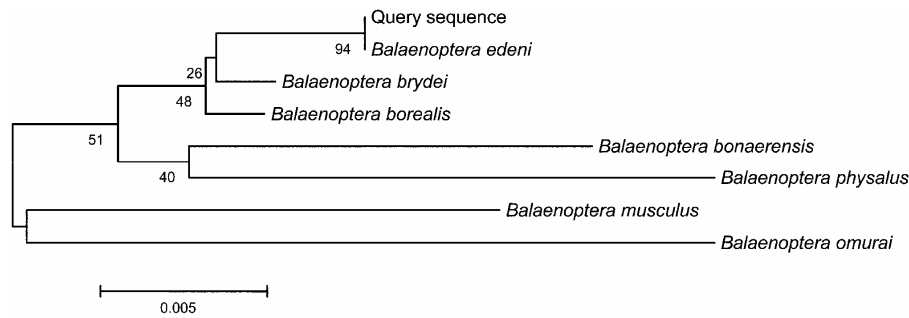


Figure 2. Neighbourhood joining tree of 16S mtDNA gene partial sequence of *B. edeni* based on reference sequences in GenBank. Numbers on the tree branches indicate bootstrap values.

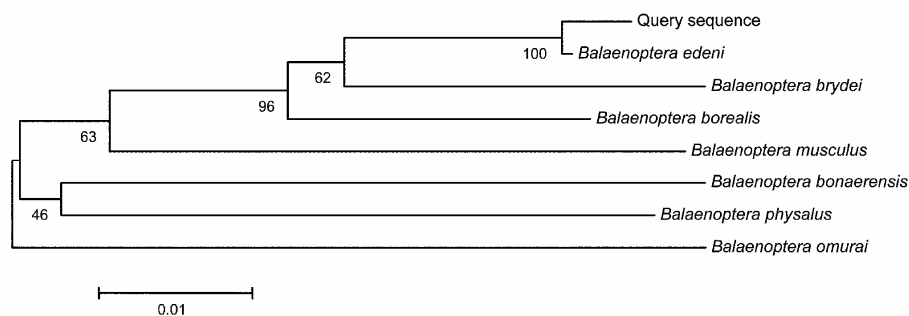


Figure 3. Neighbourhood joining tree of COI mtDNA gene partial sequence of *B. edeni* based on reference sequences in GenBank. Numbers on the tree branches indicate bootstrap values.

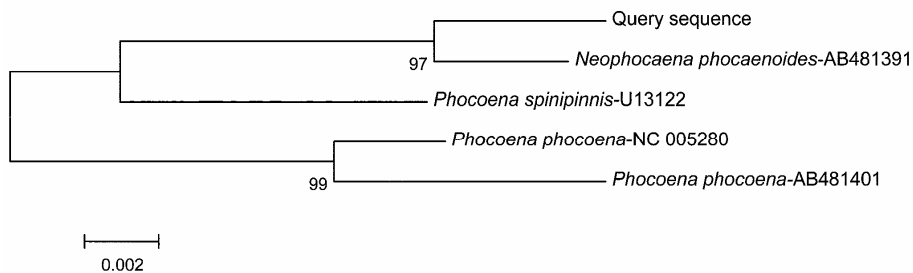


Figure 4. Neighbourhood joining tree of 16S mtDNA gene partial sequence of *N. phocaenoides* based on reference sequences in GenBank. Numbers on the tree branches indicate bootstrap values.

southwest coast of India. The earlier stranding that occurred along the southwest coast of India²¹ was from Beypore near Calicut, Kerala in 1979.

Three species have been described in the 'Bryde's whale complex', including *B. edeni*, *Balaenoptera brydei*, and *Balaenoptera omurai*, and taxonomic refinement based on mtDNA studies separated *B. brydei* (Bryde's whale) and *B. edeni* (Eden's whale) into two distinct species^{10,11}. Considering the earlier molecular taxonomic study of *B. edeni* from the east coast of India¹³ and our study, it may be inferred that the populations inhabiting the seas around India could be the 'ordinary' form of species in the Bryde's whale complex. We propose that the common name of the whale inhabiting the seas around India could be considered as Eden's whale, rather than as Bryde's whale, pending taxonomic uncertainties of the species involved in the complex. Our phylogenetic stud-

ies also corroborate the contention of related studies¹¹ that *B. edeni* constitutes a sister taxon to *B. brydei*. Further investigation is warranted to ascertain the population genetics of Bryde's whale complex inhabiting the seas around India, since recent studies recorded that gene flow between Bryde's whale populations is low and that effective management actions should treat them as separate entities to ensure continued existence of the species²².

Classified as vulnerable in IUCN red data book, *N. phocaenoides* inhabits shallow coastal waters and has the most interaction with humans and fishing gears^{1,2}. There is need to identify the sub-species of *N. phocaenoides* and their molecular data collection from the Indian waters. Frequent monitoring of the stranded marine mammals and molecular taxonomic studies would throw more light on the lesser known marine mammal diversity of the Indian coast. The present study proposes that though

osteological details would provide clues for identification of marine mammals, mtDNA sequences can be used as a reliable method for assigning species status to marine mammals stranded in putrefied condition or even not as whole organism. Further, molecular taxonomy could also be used for documenting the unknown marine mammals that often form bycatch of modern fishing gears.

1. Sathasivam, K., Status of the marine mammals of India. *J. Bombay Nat. Hist. Soc.*, 2006, **103**, 2–3.
2. Kumaran, P. L., Marine mammal research in India – a review and critique of the methods. *Curr. Sci.*, 2002, **83**, 1210–1220.
3. Simon, C., Friati, F., Beckenbach, A., Crespi, B., Liu, H. and Flook, P., Evolution, weighting and phylogenetic utility of mitochondrial gene sequences and a compilation of conserved polymerase chain reaction primers. *Ann. Entomol. Soc. Am.*, 1994, **87**, 651–686.
4. Hebert, P. D. N., Cywinska, A., Ball, S. L. and deWaard, J. R., Biological identifications through DNA barcodes. *Proc. R. Soc. London, B*, 2003, **270**, 313–322.
5. Schander, S. and Willassen, E., What can DNA barcoding do for marine biology? *Mar. Biol. Res.*, 2005, **1**, 79–83.
6. Palumbi, S. R. and Cipriano, F., Species identification using genetic tools: the value of nuclear and mitochondrial gene sequences in whale conservation. *J. Hered.*, 1998, **89**, 459–464.
7. Dizon, A., Baker, S., Cipriano, F., Lento, G., Palsboll, P. and Reeves, R., Molecular genetic identification of whales, dolphins and porpoises. In Proceedings of a Workshop on the Forensic use of Molecular Techniques to Identify Wildlife Products in the Marketplace. NOAA Technical Memoir., 2000, vol. 286, pp. 1–51.
8. Baker, C. S., Cipriano, F. and Palumbi, S. R., Molecular genetic identification of whale and dolphin products from commercial markets in Korea and Japan. *Mol. Ecol.*, 1996, **5**, 671–685.
9. Dalebout, M. L., Mead, J. G., Baker, C. S., Baker, A. N. and Van Helden, A., A new species of beaked whale *Mesoplodon perrini* sp. n. (Cetacea: Ziphiidae) discovered through phylogenetic analyses of mitochondrial DNA sequences. *Mar. Mamm. Sci.*, 2002, **18**, 577–608.
10. Wada, S., Oishi, M. and Yamada, T. K., A newly discovered species of living baleen whale. *Nature*, 2003, **426**, 278–281.
11. Sasaki, T., Nikaido, M., Wada, S., Yamada, T. K., Cao, Y., Hasegawa, M. and Okada, N., *Balaenoptera omurai* is a newly discovered baleen whale that represents an ancient evolutionary lineage. *Mol. Phylogenet. Evol.*, 2006, **41**, 40–52.
12. Dalebout, M. L., van Waerebeek, K., van Helden, A. and Baker, C. S., Molecular genetic identification of southern hemisphere beaked whales (Cetacea: Ziphiidae). *Mol. Ecol.*, 1998, **7**, 687–694.
13. Jayasankar, P., Anoop, B., Peter, R., Afsal, V. V. and Rajagopalan, M., Species of a whale and an unknown fish sample identified using molecular taxonomy. *Indian J. Fish.*, 2007, **54**, 339–343.
14. Jayasankar, P. *et al.*, Molecular identification of delphinids and finless porpoise (Cetacea) from the Arabian Sea and Bay of Bengal. *Zootaxa*, 2008, **1853**, 57–67.
15. Jayasankar, P. *et al.*, Indian efforts on the inventorization of marine mammal species for their conservation and management. *Asian Fish. Sci.*, 2009, **54**, 339–343.
16. Jefferson, T. A., Leatherwood, S. and Webber, M. A., *FAO Species Identification Guide: Marine Mammals of the World*, FAO, Rome, 1993, p. 320.
17. Palumbi, S., Martin, A., Romano, S., McMillan, W. O., Stice, L. and Grabowski, G., *The Simple Fool's Guide to PCR*, Version 2.0, Department of Zoology and Kewalo Marine Laboratory, University of Hawaii, Honolulu, 1991.
18. Folmer, O., Black, M., Hoeh, W., Lutz, R. and Vrijenhoek, R., DNA primers for amplification of mitochondrial cytochrome c oxidase subunit I from diverse metazoan invertebrates. *Mol. Mar. Biol. Biotechnol.*, 1994, **3**, 294–299.
19. Kimura, M., A simple method for estimating evolutionary rates of base substitutions through comparative studies of nucleotide sequences. *J. Mol. Evol.*, 1980, **16**, 111–120.
20. Kato, H., Bryde's whales. In *Encyclopaedia of Marine Mammals* (eds Perrin, W. F., Würsig, B. and Hewissen, J. G. M.), Academic Press, San Diego, 2002, pp. 171–177.
21. Lal Mohan, R. S., Observations on the whales *Balaenoptera edeni*, *B. musculus* and *Megaptera novaeangliae* washed ashore along the Indian coast with a note on their osteology. *J. Mar. Biol. Ass. India*, 1992, **34**, 253–255.
22. Kanda, N., Goto, M., Kato, H., McPhee, M. and Pastene, L., Population genetic structure of Bryde's whales (*Balaenoptera brydei*) at the inter-oceanic and trans-equatorial levels. *Conserv. Genet.*, 2007, **8**, 853–864.

ACKNOWLEDGEMENTS. We thank Dr Radhakrishna Pillai, Director, Rajiv Gandhi Centre for Biotechnology, Thiruvananthapuram for support. K.M. and A.B. thank Kerala State Council for Science, Technology and Environment for financial support.

Received 14 September 2009; revised accepted 12 October 2010