TABLE I
Ascorbic acid content of some freshwater fishes

Name of fish (Number of samples)		Weight range Kg.	Ascorbic acid mg./100 g. (Mean and ranges)			Mg./
			Muscle	Gonad	Liver	– gland pituitary
Macrones aor (8)		1.0-3.75	1.545	4-173	4.173	0.272*
Barbus dubius (6)	• •	1.5-3.50	(0.525 - 3.642) 1.434	(1.164-9.568) 1.726	(0.622 - 7.92) 2.272	(0.04 - 0.446) 0.126
Labeo fimbriatus (4)		$2 \cdot 0 - 5 \cdot 0$	$egin{pmatrix} (0.564-2.553) \ 3.369 \ \end{bmatrix}$	(1.007 - 2.975) 4.833	(0·870~4·433) 2·396	(0·022-0·249) 0·103
Wallago attu (2)	• •	2.0-3.75	$egin{pmatrix} (0 \!\cdot\! 79 & -6 \!\cdot\! 255) \ 2 \!\cdot\! 225 \end{smallmatrix}$	$egin{array}{c} (3\!\cdot\!498\!\!-\!6\!\cdot\!95) \ 9\!\cdot\!290 \end{array}$	(1-900-2-580) 3-493	$(0.02 - 0.238) \\ 0.028$
Labeo calbasu (1)		3.0	$egin{pmatrix} (1 \!\cdot\! 880 \!-\! 2 \!\cdot\! 571) \ 0 \!\cdot\! 930 \end{smallmatrix}$	(4.879 - 13.700) - 2.130	$\frac{(2 \cdot 115 - 4 \cdot 170)}{3 \cdot 210}$	(0.023 - 0.033) 0.020
Catla catla (1) Crocodile (1)	••	$2 \cdot 0$ $200 \cdot 0$	1 · 440 2 · 836	$\begin{array}{c} 2 \cdot 750 \\ 0 \cdot 696 \end{array}$	4 · 5 40 3 · 13 4	0.030

^{*} In terms of percentage, the AA content of one of the glands was 2.35 mg./100 mg. gland.

and Ambuja Bai and Kalyani.² Pituitary glands of Macrones aor and Barbus dubius had higher ascorbic acid than that of other three fishes studied. A decrease occurs in the ascorbic acid content of the pituitary of M. aor from February through March to June, i.e., post-spawning period. Fontaine³ noted that in salmon, the ascorbic acid in pituitary gland decreases with the muturity of the fish.

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RECORD OF APOLLODOTUS PRAEFEC-TUS DISTANT (HETEROPTERA: MIRIDAE), PREDACIOUS ON STEPHANITIS TYPICUS DISTANT (HETEROPTERA: TINGIDAE), A PEST OF COCONUT PALM

MATHEN³ described the life-history and pest habits of Stephanitis typicus D. on coconut foliage. Shanta et al. reported on its additional role as a carrier of the pathogenic principle, perhaps a virus, involved in the root (wilt)

disease of coconut, a challenging threat to its cultivation in Kerala. Since then, investigations on the various aspects of the pest like seasonal abundance, vector-virus relationship and control engaged greater attention of research workers at this research station. An interesting observation was the occurrence in the field of nymphs and adults of a Mirid bug in association with populations of Stephanitis typicus D. This has been identified as Apollodotus præfectus Distant. In its record from Pusa (Lefroy) and Ceylon (Green), no description is available on its habitat. Hoffmann made mention about a Mirid bug observed attacking these lace bugs on banana in Nanning.2 Probably it is the same or nearly related. Preliminary observations on coconut seedlings at this research station by the authors showed that the swift-moving, milkwhite predatory nymphs were available in large numbers between the post-north-east and presouth-west monsoon months of December to May. In the laboratory, under caged conditions, the nymphs were feeding well on nymphs of all stages of the pest at the rate of one to seventeen host nymphs per predator nymph per day. The nymphs were also observed to suck adult lace bugs. The authors are grateful to Dr. M. G. Ramdas Menon, Indian Agricultural Research Institute, New Delhi, for identification of the insect.

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