

CHEMICAL EXAMINATION OF *AMARANTHUS SPINOSUS* LINN.

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ABSTRACT

The petroleum-ether extract of *Amaranthus spinosus* L. on application of chromatographic techniques afforded *n*-alkanes C_{28} – C_{32} and iso (-2-methyl)-alkanes C_{29} – C_{33} ; esters: transesterified and resolved into acids C_{18} – C_{32} (as methylesters) and free alcohols C_{10} – C_{26} ; aliphatic alcohols C_{10} – C_{32} ; sterols (β -sitosterol, stigmasterol, campesterol and cholesterol) and free acids C_4 – C_{32} (C_{18} group contains stearic acid, oleic acid and linoleic acid). The free amino acids identified are leucine, valine, tryptophan, alanine and serine in the aqueous ethanolic extract.

A *AMARANTHUS SPINOSUS* LINN. (Amaranthaceae) is an important medicinal plant growing wild locally and used to cure various diseases. The ash of the leaves is used in dyeing and the plant as a whole is used in snake-bite and as fodder¹⁻². The leaves of the plant contains oxalic acid³ and sugar⁴ while hemagglutins⁵ and betacyanin⁶ are present in seeds. In view of the important medicinal properties of the herb and the fact that almost no systematic work has been done so far, a chemical examination of this herb has been undertaken, the results are presented in this paper.

EXPERIMENTAL

Air-dried and powdered plant material (2 kg) collected from surroundings of Aligarh District (India) was extracted thrice with 15 lit. distilled light petroleum (60–80°) at its boiling temperature, which after removal of the solvent under reduced pressure gave a semi-solid mass (1.18%). This extract was separated into acidic (25.2%) and neutral parts (65.4%) by washing with KOH solution (10%). The purified neutral fraction (15 g) was taken in benzene and subjected to chromatographic separation over alumina (~ 400 g) and two major fractions were collected by eluting the column with petroleum-ether (60–80°) and benzene.

Petroleum-ether fraction (4.2 g) on crystallisation from acetone-alcohol yielded a colourless waxy solid (400 mg), m.p. 62–63 C°. It showed two spots on T.L.C. (silica gel-2% AgNO₃) and therefore was resolved on a column of silica gel into two parts. Hydrocarbon fraction was eluted with hexane and the ester fraction with chloroform. The separated portions were further analysed by G.L.C. [Gas chromatography was done on PYE series 104 Chromatograph Model 124 with flame ionisation detectors, two column packed with 3% SE 30 on Gas Chrom Z at programmed temperature 150–250°C (2°C min⁻¹). For identification of homologues the graphical method was employed⁷. The comparison of the retention values with those of standards was used for identification of the gas chromatographic peaks. For a quantitative evaluation areas under the peaks have been calculated.]

Hydrocarbons form two homologous series. One of the series consists of *n*-alkanes (C_{23} – C_{33}) with maximum occurrence of *n*-hentriacontane (C_{31}) (18.1%), *n*-nonacosane (C_{29}) (15.4%), *n*-heptacosane (C_{27}) (9.0%), *n*-octacosane (C_{28}) (8.3%), *n*-triacontane (C_{30}) (8.0%), *n*-dotriacontane (C_{32}) (4.9%), *n*-tritriacontane (C_{33}) (4.5%) and *n*-hexacosane (C_{26}) (3.9%) while *n*-tricosane (C_{23}), *n*-tetracosane (C_{24}) and *n*-pentacosane (C_{25}) were in traces. The other series consists of iso (= 2-methyl)-alkanes (C_{29} – C_{33}) with a maximum of C_{31} (21.1%), C_{29} (3.4%) and C_{33} (2.1%) while C_{30} and C_{32} were in traces; the structure of 2-methyl octacosane (C_{29}) and 2-methyl triacontane (C_{31}) in this series was established by G.L.C. combined with mass spectrum. Odd numbered members were prevalent⁸⁻⁹ in both the series.

Ester fraction showed characteristic i.r. bands at 1176, 1727 cm⁻¹ and was transesterified with methanol (+HCl) and the resulting products were gas chromatographed (as methyl esters of the acid and free alcohols). The series of *n*-acids (C_{18} – C_{32} in the form of methyl esters) contained mainly C_{30} (47.7%), C_{28} (28.9%), C_{32} (8.6%); C_{18} , C_{20} , C_{22} , C_{24} and C_{29} were in small amounts (1.0–3.0%) while C_{19} , C_{21} , C_{23} , C_{25} , C_{27} and C_{31} were in traces. The even numbered members were found to be predominating. The series of *n*-alcohols (C_{20} , C_{22} , C_{24} , C_{26}) appeared in traces.

Benzene eluate (8.5 g) was further chromatographed over alumina (~ 250 g) and yielded two products.

Alcohols.—Elution with light petroleum-benzene (60:40) gave the alcohol fraction. It exhibited bands at 1050, 3400 (OH) and 715, 725 cm⁻¹ ($(CH_2)_n$ in i.r. spectrum (KBr phase). When analysed by G.L.C. it gave a series of aliphatic alcohols C_{10} – C_{34} ; even numbered members were predominating, with maximum occurrence of C_{28} (45.3%), C_{30} (19.1%), C_{22} (11.6%), C_{32} (9.6%), C_{26} (6.4%) and C_{24} (2.9%) while C_{14} , C_{24} were present in small amounts and other members were in traces. However it is interesting to mention that even numbered members were not prevalent between C_{14} – C_{34} in the same series. Branched chain alcohols were also present in minute amounts.

From this mixture *n*-octacosanol (C_{28}), m.p. $83-84^{\circ}C$ (300 mg) (lit.¹⁰ m.p. $83.4^{\circ}C$) could only be crystallised from methanol (T.L.C.-homogeneous) and confirmed by preparation of its acetate¹¹ m.p. $75-76^{\circ}C$.

Sterols.—On further elution with light petroleum benzene (20:80) the fraction of sterols (3.4 g) was obtained. It gave positive Liebermann-Burchard test and yellow colour with TNM. Repeated crystallisation from ethanol afforded shining crystals (350 mg) m.p. $158-69^{\circ}C$ (T.L.C.-homogeneous). The i.r. spectrum of the sterol exhibited peaks at 3410 (OH) and 1610 cm^{-1} ($C=C$). It showed depression in m.p. when mixed with authentic specimen of β -sitosterol and gave different R_f values (Co-T.L.C.). The crystalline sterol yielded its acetate m.p. $173^{\circ}C$ ($\nu_{\text{max}}^{\text{KBr}}$ $1730, 1245\text{ cm}^{-1}$) and benzoate m.p. $185-86^{\circ}C$. G.L.C. analysis of the product indicated it to be a mixture of β -sitosterol (83.0%), stigmasterol (12.5%) campesterol (2.9%) and cholesterol (1.6%). It is worth mentioning that the presence of cholesterol is a unique finding in a lower plant (herb).

Acids.—The acidic fraction (2.2 g) was subjected to chromatographic purification over silica gel (~ 80 g). Elution with light petroleum-benzene (1:3) afforded a colourless waxy substance (330 mg) m.p. $78-81^{\circ}C$ and showed peaks in i.r. spectrum (KBr) at 718, 728 (CH_2); 1710 (CO); 2850 and 2920 cm^{-1} (CH). T.L.C. indicated it to be a mixture which could not be resolved further even by repeated column chromatography. This product on G.L.C. analysis (after esterification) represented a series of acids C_4-C_{32} with C_{16} (27.5%), C_{18} (17.2%), C_{20} (15.5%), C_{22} (10.7%), C_{24} (9.8%), C_{26} (6.0%), C_{28} (6.1%) and C_{30} (2.5%) while the other members were found in traces. C_{18} group contained saturated and unsaturated acids and was further analysed by G.L.C. (5% diethyleneglycol succinate, at $140^{\circ}C$ on polar phase) which indicated it to be a mixture of stearic acid (32.6%), oleic acid (52.6%) and linoleic acid (14.7%). Branched chain acids were also present in minute amounts.

Aqueous ethanolic extract.—The defatted plant material was also extracted with aqueous ethanol (1:1) at its boiling temperature and concentrated *in vacuo*. A dark brown syrupy liquid was obtained which showed the presence of amino acids. The concentrated extract was extracted with chloroform to remove colouring matter. Descending paper chromatography^{8,12} (Whatman No. 1) of the aqueous alcoholic extract in *n*-butanol : acetic acid : water

(4:1:5, v/v organic layer) indicated the presence of leucine, valine, tryptophan, alanine and serine (when sprayed with ninhydrin 0.1% acetone) as free amino-acids and confirmed by direct comparison with their respective reference samples.

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