from 2 N HCl and analysed again by descending chromatography (Bu: Ac: H₂O). Standard S-methyl and S-propyl cysteine sulfoxides and cycloallun were used as reference compounds.

Results and Discussion

The amino acids obtained from the ether extract (yield 0.1 g/Kg onion) on descending chromatography gave spots corresponding to S-methyl cysteine sulfoxide ($R_t = 0.17$) and S-propyl cysteine suifoxide ($R_i = 0.35$). A third spot also appeared $(R_t = 0.63)$ which may be S-propyl cysteine¹. The amino acids obtained from the ethanolic extract (yield 0.65 g/Kg onion) on chromatography gave spots corresponding to the above two sulfoxide amino acidy and also cycloallin (R_i = 0.24). The identity of the three sulfoxide amino acids in the above preparation was further proved by two-About dimensional chromatography. twelve unidentified amino acids also appeared on the same They may be the other onion chromatogram. amino acids reported in a similar study?. By recrystallisation a mixture of the above three sulfoxide amino acids was obtained from fraction 1 [eluent from IRA-93 (OH) yield 0.3 g/Kg onion] and pure cycloalliin from fractions 2 and 3, yield 0.2 g/Kg onion.

Some of the onion sulfur compounds which, from the separated et al.3 Bandyopadhyay belong therefore extract may ether S-propyl cysteine sulfoxides S-methyl and and their degradation products reported elsewhere8. The therapeutic effects of the ether extract of onion and its sulphur compounds^{4 9-11} and also the importance of its sulfoxide amino acids with relation to sulphur metabolism¹² warrant further investigation on this vegetable.

The author acknowledges with thanks Dr. G. G. Freeman, National Vegetable Res. Lab., Wellsbourn, Warwick and Dr. E. J. Matikkala, Biochemical Institute, Helsinki, for the supply of reference compounds.

Department of Biochemistry, K. T. Augusti, University of Kerala, Trivandrum, June 4, 1976.

1. Virtanen, A. I. and Matikkala, E. J., Acta Chem Scand., 1959, 13, 1898.

2. Matikkala. E. J. and Virannen, A. I., Suomen Kemistilehti, 1957. 30 B, 219.

3. Bandyopadhyay, C., Srirangarajan, A. N. and Sreenivasan, A., J. Chromatog, 1970, 47, 400.

4. Mathew. P. T. and Augusti. K. T., Ind. Jour. Exptl. Biol., 1973, 11, 573.

5. Carson, J. F. and Wong, F. F., Jour. Org., Chem., 1961, 26, 4997.

- 6. Smith, I, In Ivor Smith (editor), Chromatographic and Electrophoretic Techniques, Vol. I. Chromatography, Williams Heinemann Medical Book Ltd., London, 2nd ed., 1960, Chapter 5, p. 82.
- 7. Matikkala, E. J. and Virtanen, A. I., Acta Chem. Scand., 1967, 21, 2891.
- 8. Johnson, A. E., Nursten, H. E. and Williams, A. A., Chem. and Ind., 1971, 1, 556.
- 9. Augusti, K. T., Benaim, M. E., Dewar, H. A. and Virden, R., Atherosclerosis, 1975, 21, 409.
- 10. and —, Clin. Chim. Acta, 1975, 60, 121. 11. —, Ind. Jour. Exptl. Biol., 1976, 14, 110.
- 12. Maw, G. A., In A. Senning (editor), Sulphur in Organic and Inorganic Chemistry.
 Vol. 2, Marcel Dekkar, Inc., New York, 1st Edn., 1972, Chapter 1, p. 115.

CHEMISTRY OF TERMINALIA SPECIES

Part XV*. Chemical Examination of T. procera Roxb. In the course of our studies on the heartwoods of Terminalia species 1-4, a sample of heartwood of T. procera Roxb. was secured from Andaman is lands. Preliminary examination showed the presence of tannins and phenolic compounds, but no triterpenes.

The hexane extract contained β -sitosterol (m.p. 136-37°, acetate m.p. 125-26°), identified by comparison with an authentic specimen. The chloroform solubles contained three components which could be separated by column chromatography on silica gel. From the analysis and by comparison with the authentic samples, they were identified as (i) ellagic acid [m.p. 360°, blood red colouration with Greismeyes's reagent⁵, ν_{-}^{Nujol} : 3570(m), 3490, 1730–1705 (br), 1625, 1585, 1540 cm⁻¹, λ_{max}^{FtOH} : 255, 366, with NaOAc 256, 278, 3556] (ii) 3, 3'-di-O-methyl ellagic acid [yield 0.03%, mp. 325-27°, blood red colouration with Greismeyer's reagent5, Nujol: 3460 (br), 1715, 1580, 1550 cm⁻¹, $\lambda_{\text{max}}^{\text{EtOH}}$: 248 (log ϵ 4.75), 372 (log ϵ 4.15) unaffected with NaOAc or AlCl₃⁶] and (iii) 3, 3', 4tri-O-methyl ellagic acid [yield 0.035%, m.p. 285-87°, green-brown-yellow colouration with Greismeyer's reagent⁵, $\nu_{\text{max}}^{\text{Nujol}}$: 3440 (br), 1720, 1580, 1550 cm⁻¹, $\lambda_{\text{rosx}}^{\text{FtOH}}$: 249 (log ϵ 4.65), 370 (log ϵ 4.05) unaffected with NaOAc or AlCl₃⁶]. Their identification was further confirmed by the preparation of tetra-O-methyl ellagic acid (m.p. 340-42°) using K_2CO_3 and dimethyl sulphate in acetone. Ellagic acid was obtained (yield 0.5%) from the acetone extracts of the heartwood.

Ellagic acid is a common constituent of a number of species of *Terminalia*. The 3,3'-di_O-methyl ellagic acid was obtained from *Euphorbia formo-sanum* Hey⁷ and it occurs as a glucoside in the

heartwood of *T. paniculata* Roth⁸. The 3, 3', 4-tri-O-methyl ellagic acid was isolated from the bark and heartwood of *Eugenia maire* A. Cunn⁹. The co-occurrence of these two methyl ethers has been found for the first time in *T. procera*.

One of the authors (A.S.) is thankful to C.S.I.R., New Delhi, for the award of a Junior Research Fellowship,

Chemistry Department, A. S. R. ANJANEYULU.
Andhra University, L. RAMACHANDRA Row.
Waltair 530 003, A. Sree.

March 29, 1976.

THE UTILITY OF MANGANESE IN LACTOSE MEDIUM TO DIFFERENTIATE RHIZOBIA FROM AGROBACTERIA

AGROBACTERIA are the most common bacterial contaminants which resemble rhizobia and are often confused with them during the routine isolation procedures. However, certain biochemical tests^{1,2} have been proposed earlier to differentiate rhizobia from many species of Agrobacterium. In this connection, Clark³ has reported that a modified Bergersen's medium4 containing lactose and 20 m.e. Mn² /1 could preferentially support the growth of all the species of Agrobacterium but exclude strains of clover and medic-rhizobia. In the present investigation, this finding has been verified extensively with tropical rhizobia of diverse origin comprising 115 isolates from Cicer arietinum, 12 from Seshania bispinosa and S. sesban, and 8 isolates of Agrobacterium sp.; the latter was isolated as contaminants during routine isolation of nodule bacteria. All the joblates were inoculated on agar slopes of the following composition: lactose, 5.0 g; KNO₃,

 $1.0 \,\mathrm{g}$; MgSO₄.7H₂O, $0.1 \,\mathrm{g}$; Na₂HPO₄ (anhydrous), $0.18 \,\mathrm{g}$; agar $12.0 \,\mathrm{g}$; FeEDTA ($0.25\% \,\mathrm{W/v}$), $10.0 \,\mathrm{ml}$; MnSO₄ ($33.5\% \,\mathrm{W/v}$), $10.0 \,\mathrm{ml}$; pH, $6.8 \,\mathrm{and}$ volume made upto $1000 \,\mathrm{ml}$ by distilled H₂O. After incubating the cultures for 14 days, at $28^{\circ} \pm 1^{\circ} \,\mathrm{C}$, observations were recorded for visible growth. Infectivity of all the isolates was tested on their homologous hosts according to the method of Wieringa and Bakhuis⁵.

It was interesting to note that all the isolates of rhizobia which nodulated their hosts could not also grow on the manganese lactose medium while the isolates of Agrobacterium showed luxuriant growth, thereby lending support to Clark's finding³ and confirming the utility of the test as a diagnostic feature to differentiate Rhizobium from Agrobacterium (Table I).

TABLE 1

Growth of rhixobia and agrobacteria on modified Bergersen's medium containing lactose and 20 m.e.

Mn²⁺/1

Strains	Isolated from	No. of strains found positive	
		Growth on lactose- manganese medium	Nodulation on homo- logous host
Cicer-rhizobia (115)	Cicer arietinum		115
Sesbania-rhizobia (10)	Sesbania		
Sesbania-rhizobia	bispinosa	• •	10
(2)	S. sesban	• •	2
Agrobacterium (5)	C. arietinum	5	• •
Agrobacterium (3)	S. bispinosa	3	

Figures in parenthesis are the number of isok tes.

Our sincere gratitude is due to Dr. N. S. Subba Rao, Head, Division of Microbiology, Indian Agricultural Research Institute, New Delhi, for his helpful discussion and critical comments.

Division of Microbiology, Y. D. GAUR. I.A.R.I., New Delhi 110 012, A. N. SUN. India, March 4, 1976.

Part XIV: Ramachandra Row, L. and Suryanarayana Murthy, P., Ind. J. Chem., 1970, 8, 1047.

^{1.} Pearson, R. S. and Brown, F. P., Commercial Timbers of India, Vol. I, Government of India, Central Publications, Calcutta, 1932.

^{2.} Ramachandra Row, L. and Ramakrishnam Raju, R. *Tetrahedron*, 1967, 23, 879; *Curr. Sci.*, 1965, 34, 177.

^{3. —} and Subba Rao, G. S. R., Tetrahedron, 1962, 18, 827.

^{4. —,} Suryanarayana Murty, P., Subba Rao, G. S. R., Sastry, C. S. P. and Rao, K. V. J., *Ind. J. Chem.*, 1970, 8, 772.

^{5.} Lederer, E., J. Chem. Soc., 1949, p. 2115.

^{6.} Jurd, L., Chem. and Ind., 1959, p. 261; J. Am. Chem. Soc., 1959, 81, 4610.

^{7.} Shinoda, H. and Kun, C. P., J. Pharm. Soc., Japan, 1931, 51, 50.

^{8.} Ramachandra Row, L. and Subba Rao, G. S. R., Tetrahedion, 1962, 18, 357.

^{9.} Briggs, L. H., Cambie, R. C., Lowry, J. B. and Seelye, R. N., J. Chem. Soc., 1961, p. 642.

Kleczkowska, J., Nutman, P. S., Skinner, F. A. and Vincent, J. M., In Identification Methods for Microbiologists, Part B. Ed. Gibbs, B. M. and Shapton, D. A., Acad. Press, London, New York, 1968, p. 51.

^{2.} Gaur, Y. D., Sen, A. N. and Subba Rao, N. S., Curr. Sci., 1973, 42, 545.

^{3.} Clark, A. G., Appl. Bact., 1969, 32, 248.

^{4.} Bergersen, F. J., Aust. J. Biol. Sci., 1961, 14, 349.

Wieringa, K. T. and Bakhuls, J. A. Plant and Soil., 1957, 8, 254.