AUSTRALOXYLON FROM THE KAMTHI BEDS OF LOWER GONDWANA, INDIA

A new species of Australoxylon, A. kanhargaoense has been described from the Kamthi Beds (Lower Gondwana) of Kanhargaon village, Chandrapur District, Maharashtra, India.

Australoxylon Marguerier, 1971

Australoxylon kanhargaoense sp. nov.

Diagnosis

Pycnoxylic secondary wood; growth rings clear (macroscopically 5-7 mm wide), late wood 14 tracheids wide (crushed), early wood tracheids polygonal to squarish. Medullary rays in cross section placed at an interval of 3-6 trachieds; rays homogeneous, 1-2 seriate, 1-30 cells deep, average height 9-10 cells (in 25 counts). Tangential surface of tracheids pitted, pitting 1-3 seriate, hexagonal, alternate and contiguous, adial wall pits c t tangentially appearing as beads. wall pitting 1-3 seriate, pits circular, alternate-opposite-subopposite, separatebordered. contiguous (mixed type of pitting), pits disposed into groups of 2, 3, 4; cross-field pits 1-7, mostly 4, bordered, sub-circular in shape

Holotype: Regd. No. 35307, Birbal Sahni Institute of

Palaeobotany, Lucknow.

Locality: Kanhargaon village, Chandrapur District,

Maharashtra, India.

Horizon: Kamthi Beds, Lower Gondwana.

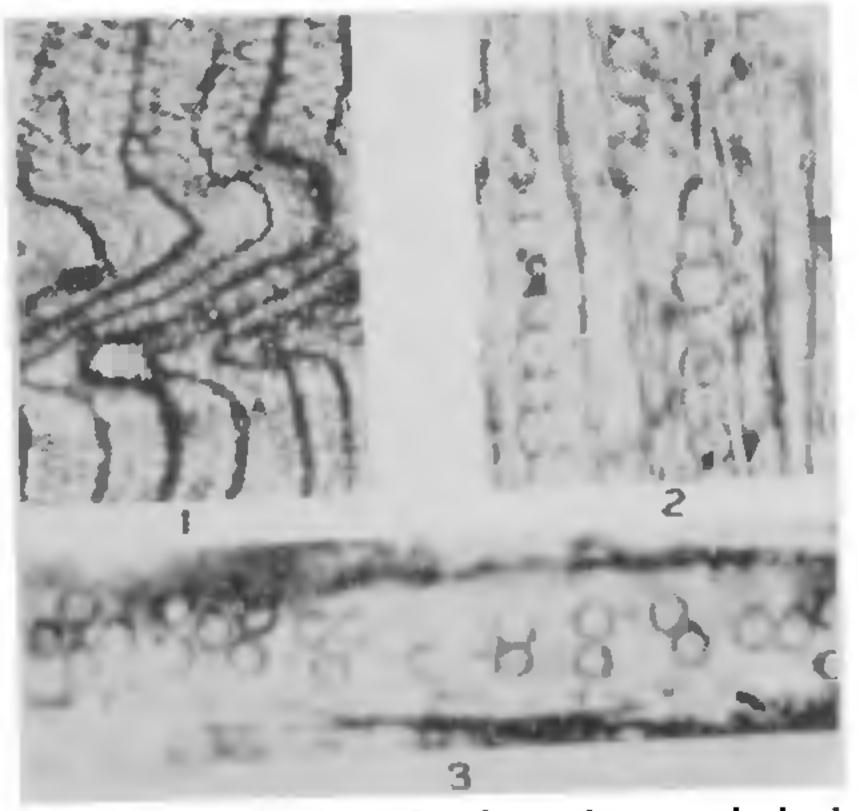


Fig. 1. Cross-section showing the crushed late wood and growth ring zone, 75

Fig. 2. Tangential longitudinal section showing the beaded appearance of tangential trachied walls, 225

Fig. 3. Radial longitudinal section showing opposite—alternate pits and groups of 2, 3, 4 pits, 500 A.

According to Marguerier³, the main diagnostic features of Australoxylon are mixed type pitting and arrangement of 2, 3, 4, or 5 pits into clusters. Marguerier described two species, viz., A. teixeirae (Type species) from the Lower Permian of Mozambique and A. natalense from the Ecca Formation of Natal. A. teixeirae differs from A. kanhargaoense in lacking tangential wall pits and A. natalense differs in having well differenciated growth rings. Hence, the present wood is referred to a new species.

It is interesting to add that mixed pitting and grouping condition of pits has also been found in *Dadoxylon* chandaensis Chitaley¹ (1949 a), *Dadoxylon* eocenum Chitaley² (1949 b), *Dadoxylon* deccani Shukla⁴ (1938), *Dadoxylon* resinosum Shukla⁵ (1944).

Birbai Sahni Institute of Palaeobotany, Lucknow, March 27, 1978.

M. N. V. Prasad. Shaila Chandra.

- 1. Chitaley, S. D., J. Indian Bot. Soc., 1949 a, 28 (3), 172.
- 2. —, Ibid., 1949 b, 28 (4), 227.
- 3. Marguerier, J., 96 Congr. nat. Soc. Savantes, Toulouse, 1971, 5, 99.
- 4. Shukla, V. B., J. Indian. Bot. Soc., 1938, 17 (5-6), 355.
- 5. —, *Ibid.*, 1944, 23 (3), 83.

INDUCED MALE STERILE LEAF MUTANTS IN CAJANUS CAJAN (L.) MILLSP.

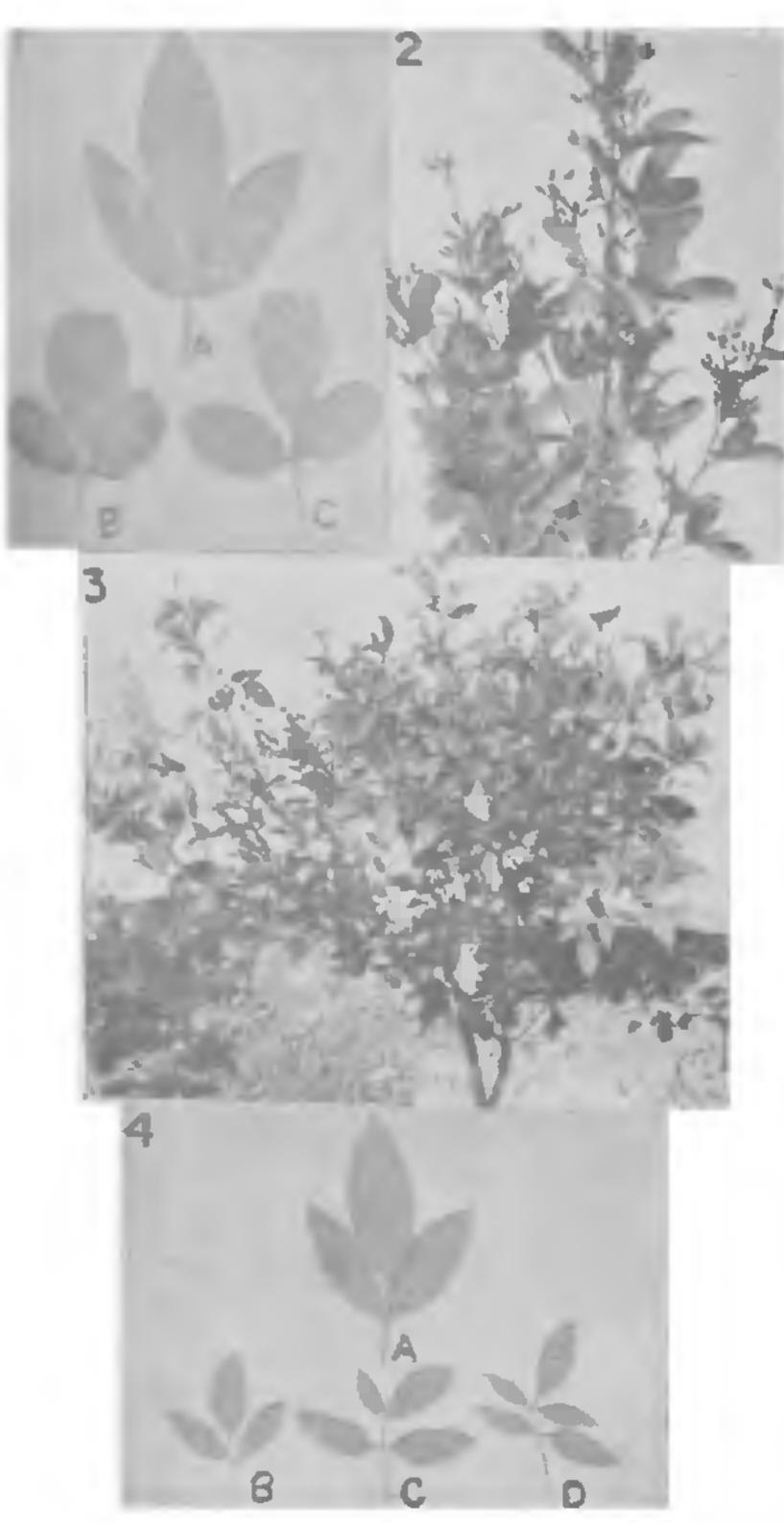
THE utilization of male sterility has become a handy tool in the proluction of hybrids, as it eliminates a laborious process of hand emasculation and pollination. Artificial induction of male sterility has been possible with the use of physical and chemical muragens¹⁻⁶. However, there are very few reports of male sterility in pigeon-pea which would promote out-crossing⁷⁻⁹. The present paper deals with the induction of male-sterile leaf mutants in an early maturing var. Pusa Ageti of Cajanus cajan (L.) Millsp. with ethyl methane sulphonate (EMS) and gamma-rays.

The seeds of Cajanus rajan obtained from Division of Genetics, I.A.R.I., New Delhi, pre-soaked for 14 hours in distilled water at room temperature, were treated with (260 seeds per treatment) 0.1, 0.2 and 0.3% aqueous solutions of EMS for 6 hours. The dry seeds were also irradiated with 10, 20 and 25 kR of gamma-rays at I.A.R.I., New Delhi. The themically treated seeds after thorough washing, and the irradiated and untreated seeds (control, were sown for obtaining M₁ generation. The seeds collected from each M₁ plant on individual plant basis were sown in the field in randomized-block-single row design.

Two types of male-sterile leaf mutants following 0.3% EMS and 25 kR of gamma-rays respectively were recorded from two individual plant progenies in M₂ generation.

LMS induced male-sterile leaf mutants

Three mutants of this type were screened. They attained a height of 85-95 cm as compared to that of control plants (115 cm). The leaflets varied from obsordate or somewhat oval in shape with obtuse ap.x. The terminal leaflet of each leaf, however, was slightly notched (Fig. 1 B & C). The flowering in these mutants was delayed by nearly a month in com-



FIGS. 1-4. Fig. 1. Leaves of control and male-sterile leaf mutant. A. Control; B. & C. EMS Induced mutant. Fig. 2. flowering branches of EMS induced male-sterile leaf mutant showing deistogamous thowers. Fig. 3. Growth habit of gamma-rays induced male-sterile multifoliate mutant. Fig. 4. Leaves of control and gamma-ray induded mutant. A. Control; B, C & D. Tri-, Tetra- and pentafoliate leaves of mutant.

parison with the controlled ones. All the buds of these mutants failed to bloom throughout their existence (Fig. 2). Two types of cliestogamous flowers were present on each mutant, viz., normal and abnormal flowers. The abnormal flowers exhibited bicarpellary condition of the ovary. In the flowers, the wing petals remained outside the standard petal, whereas the abnormal flowers invariably showed birarpellary condition of the ovary. None of these mutants produced any pod because of a very high degree of pollen sterility (97.22-99.30%) as tested by Alexander's method¹⁰.

Gamma-rays induoed male-sterile multifoliate mutant

A single male sterile multisoliate mutant was screened. It exhibited reduction in its height (80 cm) as compared to control plants (Fig. 3). Besides, normal trifoliate leaves, some tetra and pentasoliate leaves were also present on the lower branches of the mutant (Fig. 4).

The flowering in this mutant was delayed about a month as compared with the control plants. All the floral buds were cleistogamous throughout their life span, except for 5–10 buds. This mutant also failed to produce any fruit and exhibited pollen sterility between 34–57%. The causes of fruitlessness are yet to be worked out.

Sincere thanks are due to Doctors J. S. Dhakre, S. V. S. Chauhan, M. P. Singh and R. Kumar for their valuable help.

Department of Botany, R.B.S. College,

S. N. CHATURVEDI.

R. P. SHARMA.

Agra,

April 7, 1978.

- Shiv Raj, A., Rao, N. G. P., Ramana Rao, B. V. and Razvi, H. A., Indian Oil Seeds J., 1962, 6, 24.
- 2. and Ramana Rao, B. V., Ibid., 1963, 7, 156.
- 3. Hussein, H. A. S. and Disouki, I. A. M., Z. Pflanzenzuchtg., 76, 190.
- 4. Nerker, Y. S., Ph.D. Thesis, I.A.R.I., New Delhi, 1970.
- 5. Sree Ramulu, K., Proc Indian Acad. Sci., 1971, 74, 161.
- 6. —, Theor. Appl. Genet., 1972, 42, 101.
- 7. Chaturvedi, S. N. and Sharma, R. P., Curr. Sci., 1978, 47, 264.
- 8. Kaul, C. L. and Singh, S. P., Indian J. Agric. Sci., 1967, p. 37.
- 9. Reddy, B. V. S., Reddy, L. G. and Murthy R., Tropic. Grain Logume Bullstin, 1977, 7, 232.
- 10 Alexander, M. P., Stain Technol., 1969, 44, 177.