

A CALCULATOR PROGRAMME FOR FACTORIAL EXPERIMENTS

This programme can be used to analyse data from a factorial experiment in 3 factors A, B and C at levels l, m and n respectively replicated p times laid out in RBD or CRD using any machine having specifications given by Abdurazak¹. Codes have been used to identify the various numbers in the print out. The codes used are $\emptyset, 1, 2, 12, 3, 13, 23, 123$ and 11 respectively for replication, A, B, AB, C, AC, BC, ABC and error. Many characters recorded from the same experiment can be processed. Each character has to be assigned a reference number to recognise its output in the print out. The codes and reference number are printed with a negative sign. The name of the design is also to be supplied in the form of a code; \emptyset for RBD and 1 for CRD. Two factor experiments can be analysed by putting $n = 1$, and simple RBD/CRD by putting $m = 1$ and $n = 1$. If $r = 1$, the highest order interaction will be taken for error. The total number of data registers should be greater than or equal to $l + m + n + p + lm + ln + mn + 38$. All inputs are printed and spaces provided at appropriate places.

Input

(a) Parameters : Design code, p, l, m, n , Student's t at df for error. (b) Reference no. (c) The data in the order $a_1 b_1 c_1 r_1, a_1 b_1 c_1 r_2, \dots, a_1 b_1 c_2 r_1, \dots, a_1 b_2 c_1 r_1, \dots, a_2 b_1 c_1 r_1, \dots, a_2 b_m c_n r_p$.

Output

Grand total, total SS ; means for replications; marginal and two factor combination means in the order A, B, AB, C, AC, BC ; CD for these means except replications; the SS, df , MS and F for replications, A, B, AB, C, AC, BC, ABC and error respectively.

The programme is given below :

Name of the programme : FACTORIAL

000	AG4 HP T1	181	M11 F019 0 S11
007	HP S10 + 37 = S031	192	M019 HP S020
020	1 S1 S11 T0 T2	202	0 S021 11 S029
030	11 T3 21 T4 31 T9	213	G0 HA ± P M32 T2
042	R2 S0 HP T8	224	M34 T4 M36 T6
050	1 T5 10 T6 20 T7	234	0 S30 D31 S10
061	MR9 I9 + MR2 = SR9	244	R6 T9 M33 T3 M37 T7
074	I2 I3 I4 I6 I7	258	R9 T6 M35 T5
084	MR7 × 10 + R0 = SR4	267	38 T1 0 S0 = 1 T8
097	MR5 × R8 = SR2 I5	278	HP F0 FR1 × = F30 I1
109	R8 - 1 × MR6 = SR3	292	I8 M1 E R8 ≤ 278
121	M0 E R5 ≤ 061	303	M0 F31 FR2 FR3 FR4
130	I0 3 E R0 ≤ 042	317	FR5 FR6 FR7 × = F10
140	M10 S1 - 1 × S11	331	I5 I6 I7
151	(M8 - 1) = S19	337	R5 + 1 = E M36 E ≤ 267
161	M1 × M8 = S9 =	351	I3 I4 R3 + 1 =
170	0 S38 E R1 ≤ 192	360	E M34 E ≤ 258
		369	AI2 R2 + 1 =
		377	E M33 E ≤ 244
		386	M31P × = ÷ M9 = S0 ±
		399	F30 M30 PA 11 T7
		411	38T0 T1 1 T2 21 T8
		424	MR7 E 1 T5 ≤ 461
		435	0 SR0 I0 I5
		443	MR2 E R5 ≤ 435
		453	0 SR1 → 524
		461	MR8 ± P M9 ÷ MR2 = T4
		475	MR0 ÷ R4 = P
		483	C ZR0 × = FR1 I0
		494	I5 MR2 E R5 ≤ 475
		506	MR1 ÷ R4 - M0 = SR1
		519	AR7 T9
		524	I1 I2 I7 I8
		532	7 E R2 ≤ 424
		540	0 E MR7 ≤ 553
		549	R7 T9
		553	I7 19 E R7 ≤ 540
		564	M40 + M42 = ± F44
		576	M39 + M42 = ± F43
		588	M10 ÷ M1 - M0 -
		598	S46 M41 - M42 -
		609	M43 - M44 = S45
		620	M39 + M40 = ± F041
		633	0 E M19 ≤ 657
		642	M30 - M46 - M38 = S46
		657	R9 + 27 = T8
		665	MR8 ÷ MR9 = S010
		677	2T0 12T1
		684	0 E MR1 ≤ 713
		693	M10 × 2 × MR0 ÷ M9 =
		706	√ × M20 = P
		713	I0 I1 7 E R0 ≤ 684
		725	A38 T0 21 T1 11 T3
		738	0 E MR3 ≤ 769
		747	MR1 ± P
		752	MR0 P ÷ MR3 P ÷ P

763 M1Ø = PA
769 Ø SRØ IØ II I3
779 19 E R3 ≤ 738
788 AA → 213
794 Halt.

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1. Abdurazak, M. P., *Curr. Sci.*, 1980, 49, 3.
2. Panse, V. G. and Sukhatme, P. V., *Statistical Methods for Agricultural Workers*, Indian Council of Agricultural Research, New Delhi, 1961.

CHOLESTEROL AND BIOSYNTHESIS OF DIOSGENIN BY TUBER-CALLUS OF *DIOSCOREA DELTOIDEA*

SAPGENINS, including diosgenin, have been reported from tissue cultures of a number of *Dioscorea* species and some other plants¹. Cholesterol, a steroid precursor and a key intermediate in the biosynthesis of diosgenin has been shown, by using 4-C-¹⁴ and 26-C-¹⁴ cholesterol, to be incorporated in diosgenin through callus cultures of *D. deltoidea*². Effect of cholesterol on diosgenin production by seedling-callus of *D. deltoidea* has been studied in detail³. However, they³ reported a decrease in diosgenin production when the cultures were initially fed with cholesterol. On the other hand, initially fed cholesterol has been found to remarkably increase diosgenin production by callus cultures of *Trigonella foenum-graecum*⁴ and *D. floribunda*, *Costus speciosus*, *Solanum aviculare* and *S. xanthocarpum*⁵. The present authors therefore examined the effect of initially fed cholesterol on diosgenin biosynthesis by tuber-callus of *D. deltoidea*, and the results are reported in this communication.

Callus of young tuber tissue of *Dioscorea deltoidea* Wall., kept on rapidly proliferating (ca. 25-fold increase in fresh weight during 50 days) on a modified nutrient agar medium of Schenk and Hildbrandt⁶, for more than 2 years, was used as the inoculum. Composition of the culture medium in mg/l, where it differed from SH medium⁶ was: 500 NH₄NO₃, 500 meso-inositol, 2 indoleacetic acid (IAA) and 0.5 kinetin (Kn); whereas, 2,4-dichlorophenoxy-acetic acid (2,4-D) remained at the original conc. of 0.5 mg/l. Cholesterol prepared in hot ethanol was added to the medium at 50, 100, 150 and 200 mg/l conc. before autoclaving. Ten replicate cultures of each treatment were incu-

bated under 3 klx fluorescent light for 15 hr daily and at 27° ± 1° C. Sterilization procedure and other cultural conditions were as reported earlier⁷.

Callus grown on different treatments was harvested after 60 days of incubation, dried and analysed separately for diosgenin content. Each of the dried tissue samples was refluxed with 5% (v/v) HCl in 70% ethanol for 4 hr. Mixture was filtered and the residual tissue was Soxhleted with chloroform and estimated for diosgenin⁸.

The acetylated isolated compound gave mp 194°-195° C, the mmp remained undepressed and its ir spectra were superimposable with that of standard diosgenin acetate, all of which confirmed that the compound was diosgenin.

In the control, without any supplement of cholesterol, the diosgenin content was minimum being 1.03%. However, 1.03% diosgenin in tuber-callus is nearly double the content (0.682%) previously reported by Chaturvedi and Srivastava⁷ from the same tissue. This increase in diosgenin content is attributable to the composition of the changed nutrient medium. Diosgenin content of callus increased with all levels of cholesterol as compared to the control except at its lowest conc. of 50 mg/l in which the amount of diosgenin remained unchanged. Maximum diosgenin, i.e., 1.88% was biosynthesised at 100 mg/l cholesterol whereas at its higher conc. of 150 and 200 mg/l the diosgenin content of callus progressively decreased being 1.57% and 1.13%, respectively. The present results are in conformity with the reports of stimulatory effect of initially fed cholesterol on diosgenin production by callus cultures of *Trigonella*⁴ as also of *D. floribunda*, *C. speciosus* and *Solanum spp.*⁵, but not with the results of Kaul *et al.*³. Also, it is stimulatory effect of cholesterol is not affected by autoclaving (*cf.* Khanna *et al.*⁴). Addition of cholesterol at any conc. used in the present study did not inhibit callus growth to any appreciable extent. Such observations find support in the results of the studies on tissue culture of *Digitalis mertonensis*⁹ where cholesterol promoted callus growth. On the contrary, in previous investigations on callus cultures of *D. deltoidea*³ and of *Trigonella*⁴ cholesterol inhibited callus growth. That is how, such a high content of diosgenin as 2.58% obtained with the use of a combination of cholesterol and yeast extract has been reported to be nullified by the poor growth of cultures of *D. deltoidea* so much so that there has been "..... no appreciable net gain in diosgenin yield per flask"³. In the light of such observations, the 1.88% diosgenin obtained in the present study by feeding 100 mg/l cholesterol accompanied by prolific growth of callus appears to be quite significant.