the treatment. In the case of A. verrucosa one of the clones (A622c) differed from the parent (A622) and the other clones (A622a, b and d). Studies on bluegreen algae under certain culture conditions sometimes lead to highly complex taxonomic situations.

The author is grateful to Prof. C. V. Subramanian and Prof. V. S. Sundaralingam for their encouragement.

23 April 1981

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FUSARIUM SOLANI (MART.) SACC.—A NEW VASCULAR PARASITE INDUCING WILT IN MUSKMELON

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FUSARIUM wilt of muskmelon induced by one of the Fusarium spp., viz., Fusarium solani (Mart.) Sacc. is a serious disease causing severe damage to the crop. Extensive work done on the histopathological aspects of other wilt diseases has revealed many interesting findings regarding the host parasite interaction. Fusarium solani (Mart.) var. cvarum was observed to penetrate through rootlets and tracheid walls of muskmelon plants¹. Studies on comparison of Fusarium wilts of cucumber and melon, revealed that F. oxysporum led a parasitic existence in the wood vessels, whereas F. solani primarily attacked the cortex in the root and collar region². Cortical infection of F. solani was also emphasised later³.

The present study revealed that the mycelium of F. solani moved inter-and intracellularly throughout the

cortex, medullary rays and ultimately towards xylem vessels. Xylem colonization was also found in *F. solani* (figure 1). However, many of the xylem vessels were often found to be blocked with tyloses (figure 2) and a dense material. This occlusion was sometimes partial or sometimes total as regards the individual vessels. Vascular infection of *F. solani* noticed in the

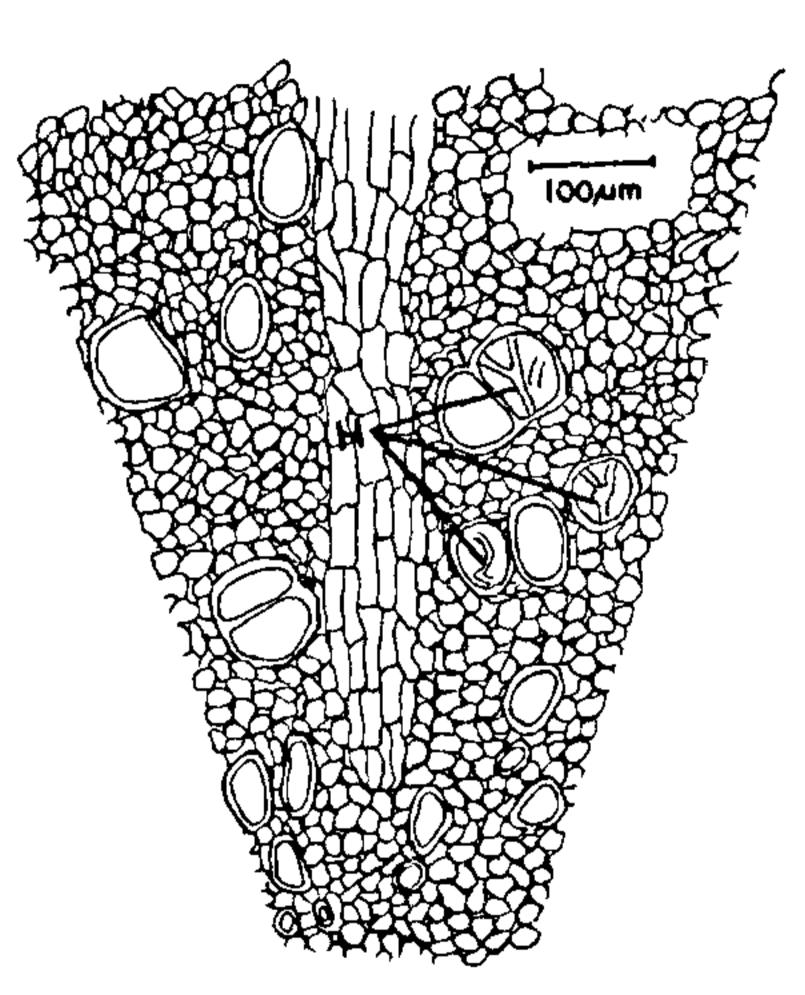


Figure 1. Portion of cross-section of Fusarium solani (Mart.) Sacc. infected muskmelon root showing hyphae (H) in xylem vessels.

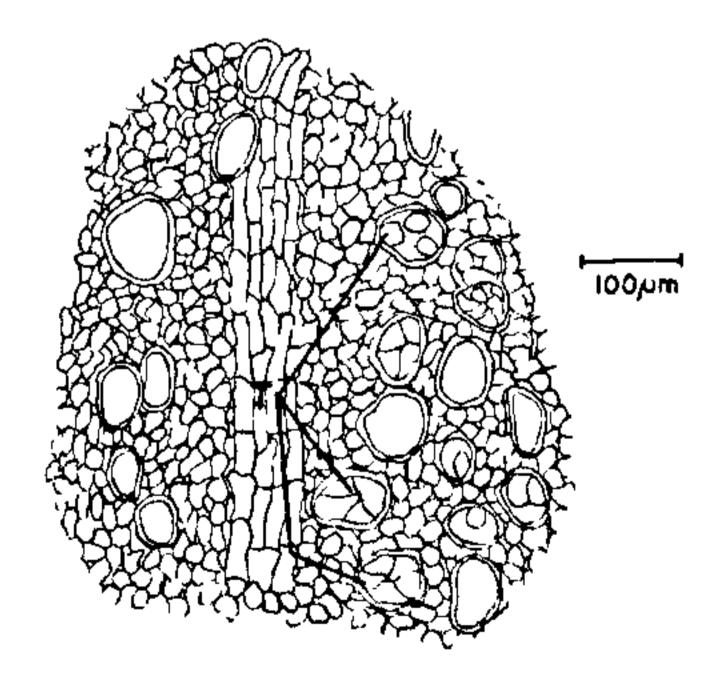


Figure 2. Portion of cross section of Fusarium solani (Mart.) Sacc. infected muskmelon root showing tyloses (T) in xylem vessels.

present investigations has thrown light into a new direction by which *F. solani* can also be called a vascular wilt pathogen, a designation which was mainly withheld by the 'Elegans' section among the *Fusaria*. This happens to be the first record of vascular colonization of this species (*F. solani*) in the roots of a

crop plant. It will perhaps be relevant to mention that work on another muskmelon isolate of *F. solani* disclosed complete absence of the usual root rotting in that particular case but the pathogen was never found in the vessels⁴.

Grateful thanks are due to Dr. L. M. Joshi, for facilities. PR acknowledges the award of a fellowship from ICAR.

22 October 1981

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INTERMEDIATE SHOOT APEX OF *PAPAVER* SPECIES

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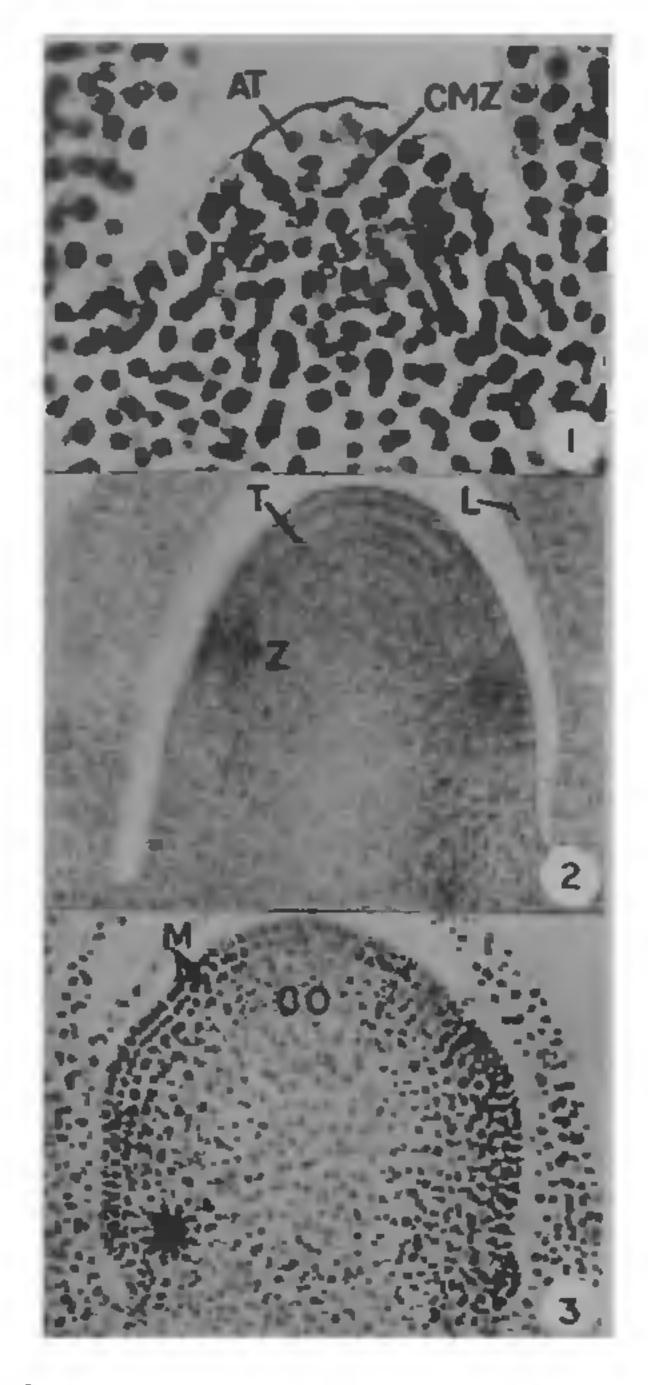
Considerable progress has been made in the last few years in anatomical studies of shoot apices as influenced by environmental factors such as photoperiod \$\frac{1}{2}\$. It was found that meristems of plants held under non-inductive conditions pass from the vegetative stage to an intermediate stage. This condition was described \$\frac{3}{2}\$, as intermediate because the apex continues to initiate new leaves but gradually acquires new characteristics that are partly vegetative and partly transitional. This paper attempts to characterise the vegetative and intermediate apices in Papaver rhoeas Linn. and P. somniferum Linn. using anatomical techniques.

The vegetative shoot apex is a low to high dome depending on the age and plastochronic stage (table 1) and shows a cytohistological zonation pattern superimposed on a tunica-corpus organisation. A gradual age-related increase in size of the apex is maintained from germination to seven weeks. The tunica in the active vegetative apex is single-layered in *P. rhoeas* and two-layered in *P. somniferum*. At the summit of the apex a few tunica cells are larger and more vacuolated, so the term axial tunica is used for these cells (figure 1).

The central mother cell zone (CMZ) at the summit of the apical dome shows a group of large, lightly stained and irregularly arranged cells. Occasional divisions in this zone contribute cells to the peripheral

TABLE 1
Average height and diameter of the shoot apex in
Papaver spp.

Plant spp. & stages	Height (in μ)	Diameter (in μ)
P. rhoeas		·
Vegetative	18-75	60-140
Intermediate	61-106	80-146
Transitional	70-119	90-164
P. somniferum		
Vegetative	20-115	40-146
Intermediate	44-180	90-160
Transitional	51-191	96-181



Figures 1-3. L. S. of the shoot apex in Papaver spp. 1. P. rhoeas showing cytohistological zonation in the vegetative apex (× 500). 2. P. somniferum showing elongate, less zonate intermediate apex (× 300). 3. P. rhoeas showing early mantle-core organisation in transitional apex. Note the lighter staining in the axially located cells (×255). (AT, axial tunica; CMZ, central mother cell zone; CO, core; L. leaf; M. mantle; PM, pith meristem; PZ peripheral zone; T, tunica; darts indicate mitotic figures).