The chromosomal aberrations scored was 6.36% with 15 min exposure and 21.25% with 60 min exposure in fixations made after 24 hr of irradiation.

The present results support the study of Sarma and Agrawal¹ on R. hieroglyphicum and O. gunnii. In all the three green algae, O. gunnii proved to be more sensitive to ultrasonic waves, showing 31.84% chromosomal aberrations at the highest dose of 60 min, R. hieroglyphicum showing 25.64% chromosomal aberrations was found more resistant that O. gunnii but sensitive in comparison to O. virceburgense, showing 21.25% of chromosomal aberrations. Thus, O. virceburgense proved to be more resistant in showing chromosomal aberrations than O. gunnii and R. hieroglyphicum. This may be attributed to the smallest chromosome size of O. virceburgense, providing comparatively less area for the action of ultrasonic waves.

25 March 1982

- 1. Sarma, Y. S. R. K. and Agrawal, S. B., The Nucleus, 1981, 24, 75.
- 2. Godward, M. B. E., New Phytol., 1942, 41, 293.
- 3. Godward, M. B. E., Nature (London), 1948, 161, 203.

A NEW LEAF SPOT DISEASE OF MAIZE FROM KUMAON, HIMALAYAS

S. K. Pant, N. P. Melkania and H. C. Joshi Vivekananda Laboratory for Hill Agiculture, ICAR, Almora 263 601, India.

DURING the survey of diseases on maize (Zea mays L.) a severe leaf spot disease was observed at Vivekananda Laboratory experimental farm, Hawalbagh (Almora). The disease made its appearance as small chlorotic spots with a light-coloured halo and 0.3 to 0.5 cm in diam. The diseased portions of the leaf were sterilized in 0.1% mercuric chloride solution and planted on potato dextrose agar medium in sterilized Petridishes. It was identified as Curvularia lunata var. aeria (Wakker) boedifn (Cochliobolus lunatus) Nelson and. Haasis) by comparing the characteristics with the type description in literature 1-3. Finally, the pathogen was confirmed from CMI, Kew, England (IMI NO. 263048a).

A perusal of literature⁴⁻⁷ indicates that the disease is prevalent only in the warm tropical part (Johner, Rajasthan) of the country. The present investigation, therefore, reveals the existence of the pathogen in temperate Himalayan region of India.

The authors are highly indebted to the Director, Vivekananda Laboratory for Hill Agriculture, Almora for providing research facilities. We are also grateful to the Director, CMI, Kew, England for confirmation of the pathogen.

12 May 1982

- 1. Barnett, H. C., Illustrated genera of imperfect fungi, (second edition), Burgress Publishing Co., Minnesota., 1960.
- 2. Ellis, M. B., Dematiaceous Hyphomycetes, CMI, Kew, Surrey, England, 1971.
- 3. Subramanian, C. V., Hyphomycetes, ICAR, New Delhi, 1971.
- 4. Bilgrami, K. S., Jamaluddin and Rizwi, M. A., Fungi of India, Part I, Today and Tomorrow's Printers and Publishers, New Delhi, 1979.
- 5. Mukerji, K. G. and Juneja, R. C., Fungi of India, (1962-72), Emkay Publications, Delhi, 1974.
- 6. Payak, M. M. and Sharma, R. C., An inventory and bibliography of maize diseases in India, IARI, New Delhi, 1980.
- 7. Sarbhoy, A. K., Varshney, J. L. and Agarwal, D. K., Fungi of India, 1971-76, Navyug Traders, New Delhi, 1980.

OCCURRENCE OF URSOLIC ACID IN EUCALYPTUS LEAVES

RAMESHWAR DAYAL Forest Research Laboratory, Bangalore 560 003, India.

No chemical work seems to have done on the eucalyptus species, and this is taken up for investigation in this paper. The leaves of *E. alba*, *E. crebra*, *E. grandis*, *E. melanophloia*, *E. microtheca*, *E. rudis*, *E. staigeriana*, *E. tesselaris* and *E. torelliana* were extracted with petroleum ether, acetone and alcohol respectively. Acetone extracts on column chromatogrphy over silica gel yielded compounds A (minor), B (major) and some minor components.

Compound A was identified as 5-hydroxy-4', 7-dimethoxy-6, 8-dimethyl flavone (Eucalyptin)¹ by NMR, IR and by direct comparison with an authentic sample.

Compound B crystallised from CHCl₁-MeOH as needles m.p. 286 88°, [α]_D + 69° (CHCl₁) and gave + ive L.B. test (Pink -> Blue). Acetate, m.p. 245-46°, [α]_D + 58° (CHCl₂). It was identified as Ursolic acid² by IR, NMR and by direct comparison with an authentic sample.

TABLE 1

Percentage of ursolic acid in the leaves of various

Eucalyptus species

Name of		% Ursolic	%
Eucalyptus species	Sc oil	acid	Eucalyptin
E. alba	0.30	1.50	
E. crebra	0.16	0.85	
E. grandis	0.30	0.60	0.010
E. melanophloia	0.10	0.92	0.014
E. microtheca	0.60	2.50	0.015
E. rudis	1.10	00.1	0.018
E. staigeriana	1.20	1.00	
E. tesselaris	0.25	1.30	0.037
E. torelliana	0.20	0.64	0.032

This is the first report on the occurrence of ursolic acid and eucalyptin (excepting in *E. torelliana*) in the respective eucalyptus species. These results also indicate that the *E. microtheca* leaves can be used for the isolation of large amounts of ursolic acid.

The petroleum ether and alcohol extracts did not give any compound.

The author is thankful to Dr. M. C. Tewari for facilities, to Dr. K. N. Subramanian, Forest Research Centre, Coimbatore for help, collecting the leaves and to Prof. J. A. Lamberton for authentic samples of acetyl ursolic acid and eucalyptin.

14 May 1982

- 1. Maria Cerecer, J., Santos, E. and Crabbe, P., Rev. Soc. Quin. Max., 1974, 18, 269.
- 2. Zurcher, A., Jeger, O. and Ruzicha, L., Helv. Chim. Acta., 1954, 37, 2145.

THE LUMINOUS BACTERIUM, BENECKEA HARVEYI IN THE GUT OF LEIOGNATHID FISHES

N. JAYABALAN* AND K. RAMAMOORTHI Centre of Advanced Study in Marine Biology, Annamalai University, Parangipettai 608 502, India.

* Present address: C.M.F.R. Institute (Annexe), Cochin 682 011, India.

Luminous bacteria are versatile heterotrophs with high adaptability. They have been reported not only from the surface waters of tropical, temperate and polar marine regions but also from a depth of several metres¹, 2. Having been known as symbionts in the light organs of fishes 3-8, saprophytes on the surface of fishes and crustaceans¹, 2, enteric bacteria in several marine organisms¹, 10, 11 and parasites in crustaceans², they also occur as general planktonic microbial populations², 12. Studies conducted in India are limited, but they are known to occur in marine, estuarine, backwater and mangrove biotopes⁸, 11. The present report is of interest as Beneckea harveyi, a species of luminous bacteria, occurs as enteric form in the gut of 6 species of leiognathid fishes (Family: Leiognathidae) viz., Gazza minuta, G. achlamys, Secutor ruconius, Leiognathus splendens, L. bindus and L. dussumieri, in Porto Novo waters.

Fish samples were collected from Porto Novo fish landing centre and the adjoining Vellar estuary (lat. 11° 30′ N, Long. 79° 46′ E) between March and May 1978. They were transferred to sterile polythene bags and brought to the laboratory in insulated containers. Totally 41 fishes (table 1) were used within 2 hr after collection. The specimens were externally sterilised with 60% ethanol to avoid contamination. Swabs of gut contents were taken using sterile cotton-tipped applicator sticks and inoculated with the sea water nutrient agar medium (SWC) consisting of 750 ml of sea water, and 250 ml of distilled water, 5g of bactopeptone, 3g of yeast extract and 3 ml of glycerol, and the pH adjusted to 7.2.

TABLE 1

Isolation of luminous bacteria from leiognathids

Species	Number of specimens utilised	Number of luminous strains isolated
Gazza minuta	7	12
G. achlamys	4	7
Secutor ruconius	8	10
Leiognathus splendens	10	18
L. bindus	7	14
L. dussumieri	5	15
Total	41	76

Cultures were grown at $25^{\circ} \pm 2^{\circ}$ C. Within 24 hr of inoculation bright luminous cotonies appeared on the medium. After 36 hr well separated single luminous colonies were picked up randomly in the dark room using sterile tooth picks and were transferred to SWC agar slants for later taxonomic characterisation.

A total of 76 luminous isolates were tested for extracellular enzymes. Especially, lipase, amylase and gelatinase are useful to differentiate *Beneckea* from the closely allied *Photobacterium*⁶. Of these, 62 iso-