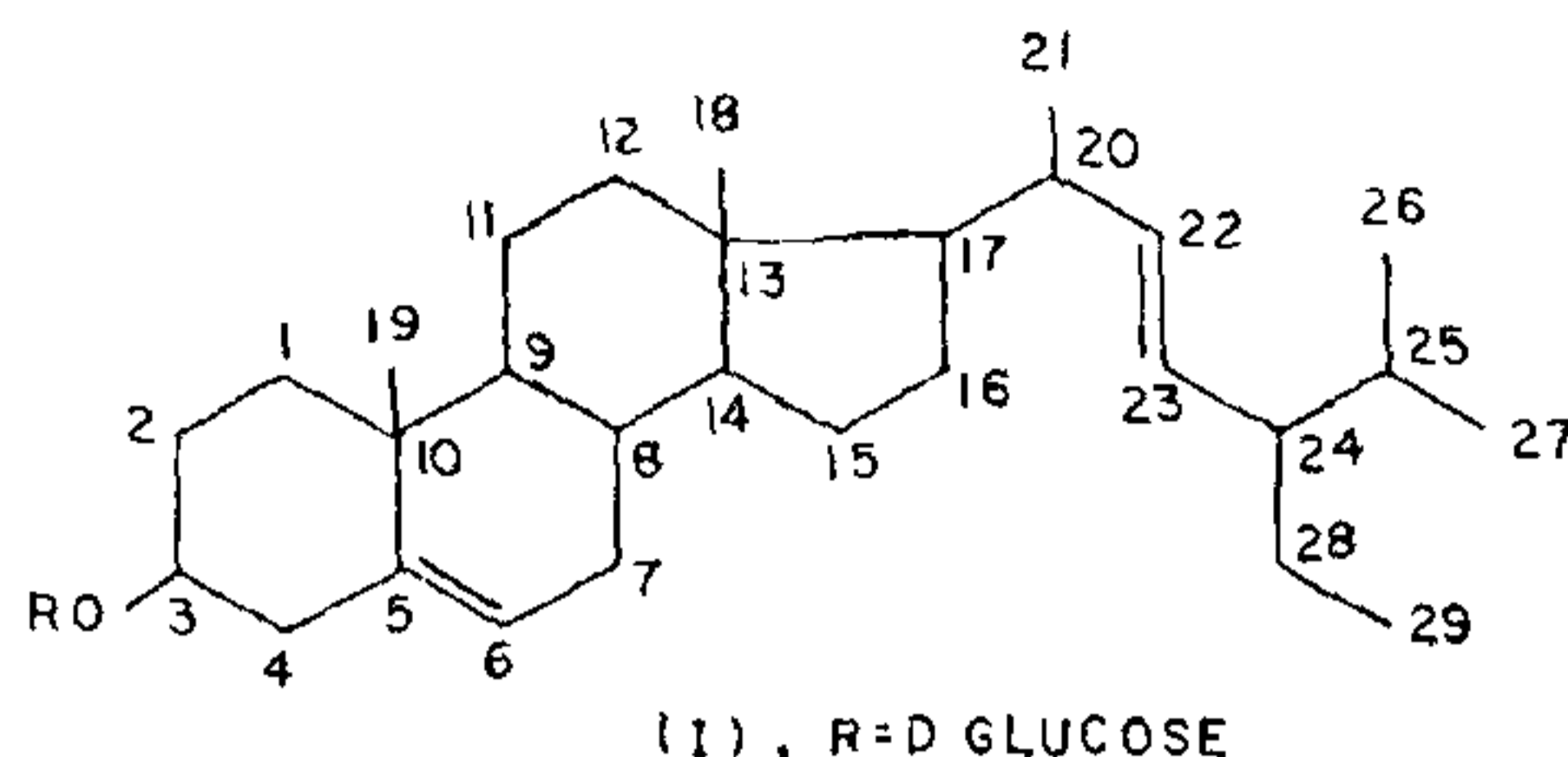


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STIGMASTA-5, 22-DIENE-3 β -O- β -D-GLUCOPYRANOSIDE, A NEW SAPONIN FROM THE ROOTS OF *SAPIUM INSIGNE*

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THE plant *Sapium insigne* is full of an acrid milk which when applied to the skin, produces vesication^{1,2}. We have examined the roots of *Sapium insigne* and isolated a new saponin; stigmasta-5, 22-diene-3 β -O- β -D-glucopyranoside (I) which is the first report in nature.

A reddish brown amorphous compound, m.p. 128–130°C (dec.) molecular formula C₃₅H₅₈O₆, (α)_D³⁰ + 16.0° (CHCl₃), gave characteristic saponin reactions: copious lather with H₂O and haemolysed red blood cells. Acid hydrolysis (7% H₂SO₄) of the saponin afforded a genin and D-glucose (co-paper chromatography and osazone formation). The genin had m.p. 153–155°, (α)_D²⁵ – 49° (CHCl₃), C₂₉H₄₈O (M⁺ at m/e 412).

From the study of IR, ¹H NMR, mass, acetate derivative oxidation³, reduction (H₂-Pt), Oppenauer oxidation^{4,5} and ozonolysis⁶ of the genin confirmed its identity as poriferasterol which was finally confirmed by co-chromatography with an authentic sample^{7,8}. Periodate oxidation⁹ consumed 2 mol of periodate with the production of 1 mol of HCO₂H per 1 mol of the saponin indicating the presence of monosaccharide in pyranose form of the sugar.

The methylated saponin¹⁰ on acid hydrolysis afforded poriferasterol (m.p., m.m.p. and Co-TLC) and 2,3,4,6-tetra-O-methyl-D-Glucose (Co-PC). The almond emulsin enzyme hydrolysis of the saponin yielded poriferasterol (m.p., m.m.p. and Co-TLC) and D-glucose confirming the β -linkage between the genin and D-glucose.

Hence the structure of the new saponin was assigned as (I)

Isolation and Purification: Air-dried and powdered roots (3 kg) of *Sapium insigne*, procured from the United Chemicals and Allied Products, Calcutta (India) were exhaustively extracted three times with ethanol (using fresh alcohol each time) under reflux for 30 days. The total ethanolic extract (30 l) was concentrated to 500 ml under reduced pressure to give a solid mass (yield 3 g) which was extracted with petroleum ether and benzene respectively. The petroleum ether extract did not yield any compound hence it was rejected. The C₆H₆ extract was purified over Al₂O₃ column (C₆H₆: CHCl₃, 1:1) and crystallized as reddish brown amorphous solid (CHCl₃) (yield 1.8 g), m. p. 128–30° (dec.); TLC (Si-gel): R_f 0.67 (C₆H₆: MeOH, 4:6) and 0.82 (CHCl₃: MeOH, 7:3), (Found; C, 73.08; H, 10.00; C₃₅H₅₈O₆ reqd., C, 73.10; H, 10.54%). The saponin (1 g) was hydrolysed with 7% H₂SO₄ (50 ml) as usual which yielded a genin and D-glucose [Co-PC and osazone, m.p. 203–4°, lit. m.p. 204–5°].

Study of the genin: (Found; C, 84.42; H, 11.62; C₂₉H₄₈O reqd; C, 84.46; H, 11.65%); IR $\nu_{\text{max}}^{\text{KBr}}$ 3400, 3050, 2950, 2870, 1640, 1630, 1425, 1390, 1060, 1050, 1000, 980, 972, 953, 837 and 800 cm⁻¹; ¹H NMR (CDCl₃, TMS, 60 MHz) δ 0.68 (d, J = 6.0 Hz, 6H, 2 \times CH₃, C-26 and C-27), 0.74 (s, 6H, 2 \times CH₃, C-18 and C-19), 0.90 (t, J = 6.0 Hz, 3H, 1 \times CH₃, C-29), 0.95 (d, J = 5.0 Hz, 3H, 1 \times CH₃, C-21), 1.2–2.00 (complex pattern, polymethylene, CH₂ and CH protons), 2.32 (s, OH), 4.48 (m, 1H, C-3), 5.12 (q, J = 4.0 and 8.0 Hz, 2H, C-22 and C-23), and 5.50 (dd, J = 4.0 and 8.0 Hz, 1H, C-6) respectively. MS: (at m/e); 412 (M⁺); 397 (M⁺ – Me); 394 (M⁺ – H₂O); 379 [M⁺ – (CH₃ + H₂O)]; 369 (M⁺ – C₃H₇); 299 (M⁺ – C₈H₁₇); 298 (M⁺ – C₈H₁₇ – H); 286 (M⁺ – C₉H₁₈); 273 (M⁺ – C₁₀H₁₉ i.e. side chain); 259 (M⁺ – C₁₁H₂₁); 257 [M⁺ – (C₁₁H₂₁ + 2H)]; 246 (M – C₁₁H₁₈O) and 231 [M⁺ – (C₁₁H₁₈O + CH₃)]; mono acetate (Ac₂O/py), m.p. 144–146° (lit. m.p. 146–47)⁷, (Found; C, 84.90; H, 11.00; C₃₁H₅₀O₂ reqd. C, 84.93, H, 11.01%); monobenzoate, m.p. 140–41°

(lit. m.p. 141°)⁷, (Found; C, 83.70; H, 10.00; C₃₆H₅₀O₂ reqd. C, 83.73; H, 10.07 %); reduction (100 mg of genin + 100 mg Pt catalyst and 20 ml glacial AcOH, shaken with H₂ for 10 hr), m.p. 142–44°, (lit. m.p. 143°)⁷, (Found, C, 83.60; H, 12.48; C₂₉H₅₂O reqd; C, 83.63; H, 12.50 %); oppenauer oxidation [100 mg of genin + 5 ml C₆H₁₂ + 120 mg (Me₂CHO)₃ Al + 10 ml Ph-Me], m.p. 76–80° (lit. m.p. 80°)⁷, (Found; C, 84.88; H, 11.20; C₂₉H₄₆O reqd; C, 84.87; H, 11.21 %); ozonolysis (100 mg of genin + 10 ml gl AcOH + O₃ ca 2%, as usual) followed by p-NO₂-Ph-NH-NH₂ treatment gave α-ethylisovaleraldehyde, m.p. 85–87° (dec.) [lit. m.p. 85–88° (dec.) and Co-TLC]⁷. (Found; C, 62.40; H, 8.00; N, 16.82; C₁₃H₂₀N₃O₂ reqd; C, 62.40; H, 8.00; N, 16.80 %).

Permethylation and hydrolysis of the saponin

The saponin (100 mg) was methylated by Hakomori's method as usual which afforded a methylated saponin, m.p. 130–32°. The methylated saponin was hydrolysed with acid (7% H₂SO₄) and worked up as usual which gave poriferasterol (m.p., m.m.p. and Co-TLC) and 2,3,4,6-tetra-O-methyl-D-glucose (RG 1.00 in *n*-BuOH: EtOH: H₂O, 5:1:4, lit. RG 1.00 in *n*-BuOH: EtOH: H₂O, 5:1:4).

Periodate oxidation of the saponin

A solution of saponin (30 mg) in ethanol (15 ml) was treated with 0.1 M NaIO₄ solution. The periodate consumed and formic acid liberated were estimated by standard procedure which corresponded to 2 and 1 mol respectively.

Enzymatic hydrolysis:

The saponin (100 mg) in ethanol (20 ml) was treated with almond enzyme solution. The liberation of D-glucose was detected by paper chromatography (Co-PC).

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CLASTIC DYKE AND THE ASSOCIATED ZEOLITE VEINS IN TALCHIR DEPOSITS, SARGUJA DISTRICT, MADHYA PRADESH

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WHILE working on Talchir deposits of Madhya Pradesh, the authors came across a clastic dyke in Gungatta river, about 15 km south of Ambikapur town. It is exposed on the left bank of the river, 1.5 km downstream from Shayam Gungatta dam site. It cuts across the sandstone and shale beds of basal Talchir Group (figure 1). The trend of the dyke is 312° with a dip about 90°. The maximum width is about 1.5 m and it extends over a distance of 15 m and both the ends are covered by soil.

The sandstone and shale beds on either side of the dyke are undisturbed and do not show any sign of deformation. The dip and strike directions of the beds are 15° NW and 225°, respectively. The dyke consists of sandy material but the grain size is finer than the associated sandstone beds.

The clastic dykes have been reported earlier from India^{1,2} and other parts of the world³⁻⁶ but the remarkable feature of this clastic dyke is the occurrence of zeolite veins penetrating throughout the body of the dyke. The major zeolite veins run parallel to the strike of the dyke but small and thin veins cut the dyke at 90°. On both sides, the dyke is bounded by thick zeolite veins. The thickness of the veins generally decreases downward. In the upper part of the dyke, the thickness is about 15 cm; however, in the lower part it