

HALOPHILA DECIPIENS OSTENFELD F. — A NEW RECORD FROM THE EASTERN COAST OF INDIA

K. K. LAKSHMANAN, S. POORNIMA and M. RAJESWARI*

Department of Botany, Bharathiar University, Coimbatore 641 046, India.

* Department of Chemical Engineering, Indian Institute of Technology, Madras 600 036, India.

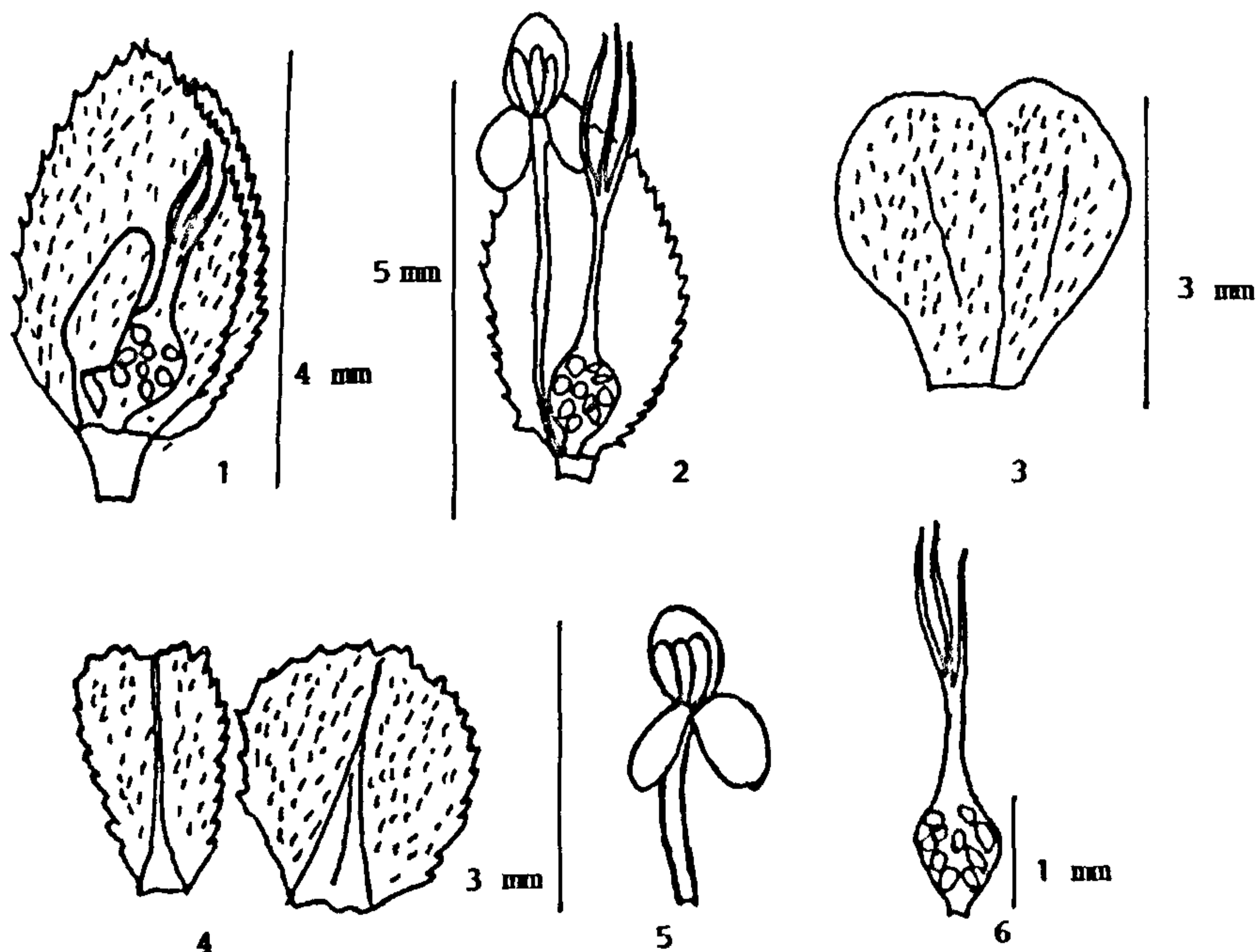
HALOPHILA DECIPIENS, the only pantropical species was collected by Dixit at Malwan near Bombay¹ (fruit 15.3.1923) and at Colaba, Bombay island (January 1928). After nearly 60 years of this record from the western coast, it is now reported for the first time from the east coast of India. The species is conspicuous in the ecosystem and in the shallow waters of Alankarathittu of Tuticorin. *H. ovata* Gaud in Freycin, *H. ovalis* (R.Br.) Hook. f., *H. stipulacea* (Forsk) Aschers and *H. beccarii* Aschers are the other species reported from Indian coasts.

Halophila decipiens Osten. f., Bot. Tidskr. 24 (1920) 260, with fig., den Hartog, Fl. Males, I 5 (1957) 410–411, f. 18; den Hartog. The seagrasses of the world, (1970) 254–258. *H. decipiens* Osten. f. var. *pubescense* den Hartog, Fl. Males I 5 (1957) 411.

The diagnostic characters of all the species of *Halophila* occurring in India are given in table 1. Specimens of *H. decipiens* can be distinguished from other species by the smaller size of the plant, presence of hairs on both surfaces of the leaf, serrulate margin, monoecious and papillae on the three stigmas (figures 1–6).

Voucher specimens are deposited in the Herbarium, Department of Botany, Bharathiar University, Coimbatore.

The authors thank the Department of Environment, New Delhi for financial assistance.



Figures 1–6. *Halophila decipiens* Ostenfeld f. 1. Young spathe; 2. Mature spathe; 3. Scale; 4. Spathe leaves; 5. Male flower; 6. Female flower.

Table 1 Diagnostic characters of species of *Halophila*

<i>H. ovalis</i>	<i>H. ovata</i>	<i>H. decipiens</i>	<i>H. stipulacea</i>	<i>H. beccarii</i>
Dioecious	Dioecious	Monoecious	Dioecious	Dioecious
Leaves 2, oblong-elliptic, surface glabrous, 1-7 cm long: 1/2-2 cm wide: margin entire: intramarginal cross veins 10-12 pairs	Leaves 2, oblong-elliptic, glabrous; 7-14 mm long: 3-5 mm wide, margin entire, intramarginal cross veins 3-11 pairs	Leaves 2, oblong-elliptic, surface rough with stiff unicellular hairs: margin finely serrulate, cross veins 4-9 pairs: 20-30 mm long and 6-10 mm wide	Leaves 2, linear to oblong, glabrous, papillose or slightly hairy 3-6 cm long: 2 1/2-8 mm wide, margin serrulate	Leaves 6-10 on erect shoot: lanceolate, glabrous margin entire, 6-13 mm long: 1-2 mm wide, cross veins absent
Flowers unisexual	Flowers unisexual	Spathe encloses one male and one female flower	Flowers unisexual	Flowers unisexual

25 May 1987; Revised 14 October 1987

1. Hartog, C. den, *The seagrasses of the world*, North-Holland, Amsterdam, 1970, p. 254.

ALTERATION OF RIBULOSE BISPHOSPHATE CARBOXYLASE/OXYGENASE RATIO BY UREA

M. C. GHILDIYAL

Department of Agronomy, University of Illinois, Urbana, Illinois 61801, USA.

Present address: Division of Plant Physiology, Indian Agricultural Research Institute, New Delhi 110 012, India.

RIBULOSE biphosphate carboxylase/oxygenase (Rubisco) catalyses carboxylation and oxygenation of RuBP to initiate photosynthesis and photorepiration respectively. Carbon dioxide and oxygen are mutually competitive substrates in these reactions¹. Modification of enzyme reactivity towards its alternate substrate might enhance plant productivity^{2,3}. The question arises whether the activity of Rubisco can be modified. Efforts have been made to seek chemical agents which modify its activity. However, most of the apparent effects were the result of artifacts in the assay procedures used^{4,5}. The present study by utilizing a sensitive assay procedure indicates that urea decreases the carboxylase/oxygenase ratio.

RuBP carboxylase/oxygenase was purified from spinach following the method of Jordan and Ogren⁴. Purified enzyme was preincubated with 10 mM NaHCO₃, 10 mM MgCl₂ with 25 mM tris (pH 8) and 0.25 mM EDTA for 1 h at 20°C. Assays were

initiated by adding 70 µg of activated protein in 25 µl to reaction mixtures at 25°C containing varying amounts (0.5, 1, 2, 5, 10 mM) of NaH¹⁴CO₃, 0.0236 mM H³RuBP, 10 mM MgCl₂, 50 mM Bicine at pH 7.85, with and without 2 M urea in a total volume of 0.5 ml. Reactions were performed in 6 ml scintillation vials which were sealed with serum stoppers. The reaction mixtures were flushed with pure oxygen for 15 min before adding NaH¹⁴CO₃ and enzyme. Reactions were terminated after 30 min by adding 0.1 ml of a solution containing 0.05 N HCl and 50 mM ZnSO₄ and stored at -15°C.

Carboxylation and oxygenation rates were determined according to Jordan and Ogren⁴. The labelled products of carboxylase reaction [(³H, ¹⁴C) glycerate-P] and the oxygenase reaction (³H-glycolate-P) were separated. Glycolate-P phosphatase was used to convert (³H) glycolate-P to (³H) glycolate which was readily separated from labelled organic phosphates by ion exchange chromatography. Radioactivity in these compounds was quantified by scintillation spectroscopy. The ratio of RuBP carboxylase/oxygenase was determined by measuring the two activities simultaneously in the same reaction mixture. This procedure eliminates the possible discrepancy when two activities were measured separately under different conditions particularly at low CO₂ concentration^{4,5}.

The ratio of carboxylase to oxygenase activity (vc/v_o) plotted against the ratio of CO₂ and O₂ concentrations present during assay is shown in figure 1. The ratio increases with increase in the ratio of the CO₂ and O₂ concentrations. However, this ratio was lower when urea was included in the assay medium.