

Detection of biogenic VOCs emitted from common tropical plant species in the Western Ghats region of India: Chamber based experiments

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Brief subhead: VOC emission from plant species in the Western Ghats

Highlights:

Experiments using a custom-made branch-enclosure to study the emission compositions of BVOCs

Measurements of BVOC emission compositions from seven dominant tropical plants in the Western Ghats of India

First BVOC emission characterization study for two plant species of *Bambusa vulgaris* and *Saraca asoca*

All selected plant species show highest emissions of isoprene followed by light alkenes

Abstract

This study, for the first time, provides the detection of biogenic volatile organic compounds (BVOCs) emission from some common plant species found in the Western Ghats of India using branch-enclosure experiments. A custom-made dynamic chamber system was deployed to collect samples from seven different plant species. Analysis of speciated BVOCs was performed using C₂-C₆ and C₆-C₁₂ VOC Analyzers to determine the emission composition and relative concentrations. Isoprene was the most abundant compound, followed by ethene, propene, α -pinene and β -pinene. Among the plant species, *Tectona grandis*, *Bambusa vulgaris* and *Psidium guajava* showed the high fractions of isoprene emissions, *Saraca asoca* showed moderate emissions, and *Manilkara zapota* and *Leucaena leucocephala* showed the lowest emissions. However, *Manilkara zapota* and *Leucaena leucocephala* showed higher emissions of both ethene and propene as compared to isoprene. This study reports the emission profiles of BVOCs from *Bambusa vulgaris* and *Saraca asoca* for the first time. This study emphasizes the importance of emission flux measurements of major plant species in different forest regions of India which is necessary to develop emission inventories of important BVOCs.

Keywords: Chamber experiment, Isoprene, Monoterpenes, Alkenes, Western Ghats, Tropical forests

1. Introduction

Emissions of biogenic-VOCs (BVOCs) including alkenes, isoprenoids (isoprene: C_5H_8 , monoterpenes: $C_{10}H_{16}$, sesquiterpenes: $C_{15}H_{24}$) and oxygenated VOCs (organic acids, aldehydes, ketones, alcohols, etc.) have a substantial impact on the global atmosphere^{1,2}. Emissions from terrestrial plants have been estimated to be a major source of BVOCs, with an annual global emission of $\sim 1150 \text{ Tg/yr}^3$. Among BVOCs, isoprene ($\sim 70\%$) and monoterpenes (11%) are the most abundant compounds emitted from the terrestrial vegetation^{1,3-6}. Among the other BVOCs, alkane and alkene ($\sim 30\%$) and oxygenated volatile species ($\sim 60\%$) have also been found to be the major compounds in some forest regions^{7,8}. These reactive trace compounds play a key role in regional atmospheric chemistry and climate^{7,9}. BVOCs are an important component of plant physiology and are involved in plant growth, reproduction and defence mechanisms¹⁰⁻¹². The emissions are sensitive to abiotic environmental factors like temperature, relative humidity, CO_2 and light intensity¹³⁻¹⁵. Therefore, the composition and fluxes of BVOCs emitted from the terrestrial plants are known to show strong species-to-specific variations^{16,17}. Studies on some tropical plant species have shown that the climate change (global warming) can bring a change in BVOC emissions in response to induced stress¹⁸.

Emissions from the tropical regions contribute more than 70% to the global budget of total BVOC^{19,20}. The South Asian region, consisting of about 15% of world's tropical forest region as of 2010²¹, has a rich diversity of tropical plant species. However, very limited information is available on their BVOC emission characteristics^{17,22-26}. Nonetheless, it is important to investigate the BVOC emissions from tropical vegetation in view of their key roles in chemistry-climate interactions. Among the South Asian nations, India has the largest geographical area with $\sim 713789 \text{ km}^2$ of area under forest cover. About 14 major forest types occupy nearly 21% of the India's total geographical area^{27,28}. The tropical-moist and tropical-dry forests cover $\sim 65\%$ of the total forest area of India²⁹. A map of the different forest types and land use/land cover of India prepared using forest type data at 5 km resolution from the Bhuvan Geo Portal (<https://bhuvan.nrsc.gov.in>)³⁰ is shown in Figure 1. Despite large forest areas, the efforts to measure the emissions of BVOCs from Indian plant species have not been systematic and comprehensive. A few studies^{17,22-26,31-35} have reported BVOC emissions from different plant species in India. The isoprene and monoterpene emission rates from common tropical plant species of the *Amarkantak-Achankmar-Biosphere Reserve* (AABR) in central India are reported in¹⁷.

During the last few decades, researchers have been trying to design and develop the branch-enclosure systems, both static and dynamic, to quantify the emissions of BVOCs from different plants³⁶⁻³⁹. Several recent advances in branch enclosures studies, using both static and dynamic enclosures, have been made. But the dynamic enclosure is more convenient and recommended for authentic measurements of BVOC emissions⁴⁰. Unlike the static enclosure, the environment and circulation can be controlled in dynamic enclosures^{11,41-43}. In addition, some studies have also shown that the changes in temperature and relative humidity inside the static enclosures. In absence of purge flow, the temperatures inside the static chamber rises, and this may affect the true estimations of BVOCs. While such uncertainties are relatively less in case of dynamic chamber-based estimations. In static chamber there is compulsion of zero air while as it is not for dynamic one. During the last few decades, the studies have been focused using the dynamic chambers in order to overcome the uncertainties associated with static chamber based studies^{41,44,45}. So far, the dynamic chamber has been the most widely used system for the measurements of BVOC emissions from the plant species⁴⁶⁻⁴⁸, and to determine its dependence on different environmental variables^{10,49-52}. However, lack of pre-established and tested protocol, and thus, inconsistency in enclosure systems and deployment methods leads to large variations in emission estimates across the globe^{53,54}. In this study, we have presented the descriptions of a custom-made chamber system and its performance during the field deployments for the measurements of BVOCs emission from the seven different plant species. The seven selected species in this study, namely: *Bambusa vulgaris*, *Saraca asoca*, *Gliricidia sepium*, *Psidium guajava*, *Tectona grandis*, *Leucaena leucocephala* and *Manilkara zapota* are few of the most common plant species found in the Western Ghats of India^{57,58}. Among these, the BVOC emission compositions from the two selected plant species (*Bambusa vulgaris* and *Saraca asoca*) have not been reported previously in the literature. Thus, this study provides a test case investigation about the composition of BVOCs emitted from these plants in the Western Ghats region. And for the remaining five species, the BVOC emission properties have been reported for other parts/regions but not for the Western Ghats. The main objectives of chamber-based experiments include detection and identification of the most dominant isoprenoid and alkene compounds emitted from the selected plant species in the Western Ghats of India.

2. Experiments and Results

2.1 Study region

The Western Ghats of India is a global biodiversity hotspot covering the states of Maharashtra, Goa, Karnataka, and Kerala^{9,55}. The geographical area of the Western Ghats is ~160,000 km², out of which ~57000 km² (35.6%) is forest cover⁵⁶. The Western Ghats forests are dominated by the moist deciduous, semi-evergreen, dry deciduous, and wet evergreen trees^{29,57}. The study was conducted in the campus of the National Institute of Oceanography (NIO) (15.5°N, 73.84°E), Goa. The area experiences typical tropical monsoon climate with the annual average precipitation of approximately 3800 mm, most of which is concentrated between the months of mid-June and September²⁷. In this region, *Peltophorum pterocarpum* (Peela gulmohar/Copper pod), *Ficus religiosa* (Peepal tree), *Ficus amplissima* (Nunurki tree), *Mangifera indica* (Mango tree), *Gliricidia sepium* (Mexican Lilac/Saranga), *Cocos nucifera* (Coconut palm), *Terminalia catappa* (Indian almond), *Saraca asoca* (Ashoka), *Tectona grandis* (Teak/Saguan), *Tamarindus indica* (English tamarind), *Leucaena leucocephala* (Kubabul), *Opuntia ficus* (Indian fig), *Psidium guajava* (Guava), *Manilkara zapota* (Chikoo), *Bambusa arundinacea* and *Bambusa vulgaris* (Bamboo) are the dominant tree species^{57,58}. Among these, we have selected seven species of *Bambusa vulgaris*, *Saraca asoca*, *Gliricidia sepium*, *Psidium guajava*, *Tectona grandis*, *Leucaena leucocephala* and *Manilkara zapota* to investigate their BVOC emission characteristics (Figure 2). These selected species were mature and not affected by any pests and diseases.

2.2 Design of custom-made chamber

We have designed a dynamic chamber for the direct sampling of BVOCs emitted from selected plant species as shown in Figure 3. The chamber is constructed from a transparent acrylic cylinder with a length of 0.60 m and a diameter of 0.25 m. The top of the chamber is closed using an acrylic plate/disk with two holes of 12 mm internal diameter (ID). These holes on the plate are used to insert the inlet and outlet lines. About 4 m long teflon tube (6 mm id) was connected to inlet for the supply of zero-air and a 50 cm long teflon tube was connected to the outlet for air sampling. The other open side of the chamber is wrapped with teflon foil after the plant-branch is inserted. The schematic diagram and onsite deployment of the chamber setup are shown in Figure 3. We collected the samples in tedlar bags (10×15 inch) made from 2 mm thick tedlar film with a capacity of 5 L (SKC Inc., catalogue no. 232-05, PA 15330-9613, USA). A polytetrafluoroethylene (PTFE) hose valve and an injection port containing a teflon fluorocarbon resin septum are attached to the bag for the sample collection and analysis without contamination or loss of species in the sampling volume.

2.3 Chamber experiments and analysis of BVOCs

The chamber experiments were performed in the daytime (between 10:30 and 15:30 hr) during 13-17 October 2022. The details of sampling and ambient conditions for each experiment are summarized in Table 1. During the experiments, we collected samples for each of the representative plant species as well as from the ambient (background) air. For each plant species, an open branch was selected for the experiments to avoid underestimation of emissions due to the shaded (non-sunlight) leaves⁵⁹. For each plant species, an exposed branch at heights of 1-2 m from the ground was selected and enclosed in the chamber. To avoid the impact of any physical stresses, the chamber enclosure was held stationary using a supporting platform. The branches with leaves were carefully placed inside the chamber to minimize the contact with the inner wall of the chamber. After enclosing the branch, the open back-end of the chamber was tightly wrapped using teflon foils. After that, a continuous zero-air (99.9999%) flow was supplied from the cylinder at a 15 psi (1034.21 hPa) (outlet pressure) i.e. slightly higher than the ambient atmospheric pressure for 20 min into the 30 L volume of the chamber. This continuous flow of zero-air into the chamber provides uniform mixing of BVOCs emitted from the branch⁴¹. The enclosure system was not completely airtight and a continuous flow of zero-air was required to avoid the inward flow of ambient air. Due to this, some pressure might be generated in the chamber, but due to the non-availability of pressure sensor, this pressure was not measured. The flow of VOC-free zero-air was preferred instead of ambient air to minimize any alteration of BVOC emissions from the plants. The samples were collected after 20 min of zero-air flow, by using a pocket pump (SKC Inc., catalogue no. 22-2301, PA 15330-9613, USA) through a teflon/silicone line extended from the outlet of the chamber. The ambient air (background) samples were collected at the location of sampled species before the start of the chamber experiment. During our experiment, we carefully inserted the branches inside the chamber to avoid any injury or breaking.

The analysis of speciated BVOCs including ethene, propene and isoprene present in the collected samples was performed using a C₂-C₆ VOC analyzer (AirmoVOC Model: A12000, Chromatotec®, Saint-Antoine, France). While monoterpene compounds including α -pinene and β -pinene were measured using a C₆-C₁₂ VOC analyzer (AirmoVOC, Model: A22022, Chromatotec®, Saint-Antoine, France). Both these instruments are based on the thermal desorption-gas chromatography coupled with a flame ionization detector (TD-GC-FID). The collected air samples were introduced into a peltier-cooled (at -15°C) adsorbent trap via a 1 m long stainless steel (SS) tube (0.25" ID). In the C₂-C₆ VOC analyzer, BVOCs samples pre-

concentrated on the adsorbent trap are desorbed into a PLOT column (Al₂O₃/Na₂SO₄, 25 m× 0.53 mm, 10 µm film thickness, Restek Corp., USA). In the C₆-C₁₂ VOC analyser, the desorbed samples were transferred into an MXT30CE column (30 m × 0.28 mm, 1 µm film thickness, Restek Corp., USA). The identification and peak area integration of different compounds were performed using VISTACHROM® software developed by Chromatotec®. Additional descriptions of VOC analyzers are provided in our previous study⁶⁰. We performed multipoint calibrations on our system using standard gas mixture (lot no: 1341032, Linde, USA) containing C₂-C₈ NMHCs at ~1 ppm with an analytical accuracy of ± 5%, and also using a permeation tube (Benzene-SN:20170725-E703, Chromatotec®, France). The calibrations were performed at five different mixing ratio values between 0-10 ppb (generated using a gas calibration unit, dynamic dilution system). The sensitivity was determined as the slope of the calibration curve and the detection limit (DL) was calculated using calibration curve through the following formula (according to guidelines of International Conference on Harmonization (ICH)).

$$DL=3.3\times\frac{\sigma}{S}$$

where, σ is the standard deviation of the y-intercept, and S is the slope of the calibration curve.

For analysis of α - and β - pinenes, system default sensitivity (response factors calculated with reference to benzene) was used. The detection limit (ppb) of ethene, propene, and isoprene were determined to be 0.167, 0.139 and 0.195, respectively.

Although many VOC compounds were detected using this analysis, we focused our study on the emissions of major BVOCs including ethene, propene, isoprene, α -pinene and β -pinene. Typical chromatograms obtained from the analysis of samples collected from the chamber experiments on different selected plant species are shown in Figure 4.

2.4 Plant specific emissions of BVOCs

Concentrations of ethene, propene, isoprene, α -pinene and β -pinene measured during the chamber experiments for seven selected plant species are given in Table 1. Among the plant species, emissions from the five plant species (*Bambusa vulgaris*, *Saraca asoca*, *Gliricidia sepium*, *Psidium guajava*, and *Tectona grandis*) show high levels of isoprene, followed by

ethene, propene, α -pinene and β -pinene (Figure 5). The sum of the concentrations of these measured BVOCs was highest for *Tectona grandis* and the lowest for *Leucaena leucocephala*. Among the plant species, *Tectona grandis*, *Bambusa vulgaris*, and *Psidium guajava* showed higher isoprene emissions; *Bambusa vulgaris* and *Saraca asoca* showed moderate emissions; and *Manilkara zapota* and *Leucaena leucocephala* showed the lowest emissions. *Manilkara zapota* and *Leucaena leucocephala* both showed high emissions of alkenes (ethene and propene) as compared to isoprene. However, all the selected plant species showed lower monoterpene emissions compared to isoprene and alkenes.

In order to have an idea about the emission profile of BVOCs, the results obtained in this study have been compared with those reported for the same or other plants in previous studies. For instance, Hakola et al. (1998)⁶¹ reported ethene, propene and 1-butene emissions from three tree species i.e., tea-leaved willow (*Salix phylicifolia*), silver birch (*Betula pendula*) and European aspen (*Populus tremula*), with highest emissions from willow. Previous studies have revealed that ethene is produced in almost all plant parts in varying concentrations and plays an important role in fruit ripening, flowering development, senescence and other physiological processes⁶²⁻⁶⁵. The emissions of isoprene and monoterpenes from some of the plant species selected in this study are also reported in previous studies^{17,24,26,31,35,66}. Similar to present study, high BVOCs emissions are reported from *Tectona grandis* in the previous studies^{22,24}. We also found significant BVOCs emission from *Psidium guajava*. On contrary, previous studies have reported lower BVOCs emissions^{17,25}.

The emission composition of BVOCs from *Bambusa vulgaris* mainly consists of isoprene. Okumura et al. (2018)⁶⁶ have reported the BVOCs emissions from 14 different bamboo species with significant fractions of isoprene from all. The BVOC emission profiles of two tropical plant species (*Bambusa vulgaris* and *Saraca asoca*) have been reported for the first time in this study. However, genus and family-level studies are available in the literature (genus-level: *Bambusa vulgaris*; family-level: *Saraca asoca*)^{17,66}. This study also revealed that the leaves of different species within the same genus are likely to exhibit similar isoprene emission characteristics. The comparison of studies for genus and family-level studies shows considerable differences in their emission characteristics^{17,66}. Okumura et al. (2018)⁶⁶ have reported *Bambusa oldhamii* and *Bambusa multiplex* as significant isoprene emitters only. Whereas in the present study, we found that *Bambusa vulgaris* also emits smaller amounts of other BVOCs (ethene, propene, isoprene, α -pinene and β -pinene). These variations in BVOCs emission characteristics could be due to the differences in their leaf structures, physiological

characteristics and genetic makeup. There could be some other reasons like different environmental conditions, use of different measurement methods, growth forms and stage, age, etc. might influence BVOCs emission and composition. Overall, the sum of measured BVOCs as well as relative composition of individual compound showed large variations from plant-to-plant. The result indicates that isoprene is the dominant BVOC in major plant species found in the Western Ghats. Consistently, previous studies also support the fact that tropical as well as broad-leaved species are mainly isoprene emitters, while coniferous or needle-like leaves are monoterpene emitters^{10,67}. A comprehensive study is required to quantify the emission fluxes of BVOCs from different tropical plant species by taking account of the seasonality in key environmental parameters.

3. Summary

A custom-made chamber (branch-enclosure) was deployed to detect the BVOC emission profiles from seven dominant plant species (*Tectona grandis*, *Bambusa vulgaris*, *Psidium guajava*, *Saraca asoca*, *Manilkara zapota*, *Leucaena leucocephala* and *Gliricidia sepium*) in the Western Ghats of India. The experiments show that the newly designed dynamic chamber is well suited for the detection of BVOCs emissions using offline TD-GC-FID and online VOCs analysers. The emission samples obtained from the branch-enclosure experiments were analysed to determine the composition and concentrations of major BVOCs including light alkenes, isoprene and monoterpenes. Two plant species (*Bambusa vulgaris* and *Saraca asoca*) were investigated for the first time for the measurements of BVOC composition. *Tectona grandis*, *Bambusa vulgaris* and *Psidium guajava* were found to be strong isoprene emitters with smaller amounts of alkenes, α -pinene and β -pinene.

Further improvements in the chamber design have been planned to measure the dependence of key environmental variables like temperature, light intensity, CO₂ concentration, and relative humidity, as they play an important role in controlling BVOC emissions. The studies of emissions from tropical tree species in the Western Ghats are important for the inventory developments and subsequent use in atmospheric chemistry modelling studies. Although this is a preliminary study in the Western Ghats, the experiments clearly highlight the potentials of BVOC emissions from major plant species and provide scope for comprehensive study in future.

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Conflict of interest: The authors declare that there is no conflict of interest.

References

1. Fiore, A. M., *et al.*, Global air quality and climate. *Chem. Soc. Rev.*, 2012, **41**, 6663-6683.
2. Kesselmeier, J. and Staudt, M., Biogenic volatile organic compounds (VOC): an overview on emission, physiology and ecology. *J. Atmos. Chem.*, 1999, **33**(1), 23-88.
3. Sindelarova, K., *et al.*, Global data set of biogenic VOC emissions calculated by the MEGAN model over the last 30 years. *Atmos. Chem. Phys.*, 2014, **14**(17), 9317-9341.
4. Llusia, J., Llorens, L., Bernal, M., Verdaguer, D. and Penuelas, J., Effects of UV radiation and water limitation on the volatile terpene emission rates, photosynthesis rates, and stomatal conductance in four Mediterranean species. *Acta physiologiae plantarum*, 2012, **34**(2), 757-769.
5. Pokorska, O., *et al.*, Emissions of biogenic volatile organic compounds from *Fraxinus excelsior* and *Quercus robur* under ambient conditions in Flanders (Belgium). *Int. J. Environ. Anal. Chem.*, 2012, **92**(15), 1729-1741.
6. Singh, A. P., Singh, R., Mina, U., Singh, M. P. and Varshney, C. K., Emissions of monoterpene from tropical Indian plant species and assessment of VOC emission from the forest of Haryana state. *Atmos. Pollut. Res.*, 2011, **2**(1), 72-79.
7. Rhew, R. C., *et al.*, Ethene, propene, butene and isoprene emissions from a ponderosa pine forest measured by Relaxed Eddy Accumulation. *Atmos. Chem. Phys.*, 2017, **17**, 13417-13438. doi: 10.5194/acp-17-13417-2017.
8. Halliday, H. S., Thompson, A. M., Kollonige, D. W. and Martins, D. K., Reactivity and temporal variability of volatile organic compounds in the Baltimore/DC region in July 2011. *J. Atmos. Chem.*, 2015, **72**, 197-213. doi: 10.1007/s10874-015-9306-4.
9. Tripathi, N., Sahu, L. K., Patel, K., Kumar, A. and Yadav, R., Ambient air characteristics of biogenic volatile organic compounds at a tropical evergreen forest site in Central

- Western Ghats of India. *J. Atmos. Chem.*, 2021, **78**, 139–159.
<https://doi.org/10.1007/s10874-021-09415-y>.
10. Aydin, Y. M., *et al.*, Biogenic volatile organic compound (BVOC) emissions from forested areas in Turkey: Determination of specific emission rates for thirty-one tree species. *Sci. Total Environ.*, 2014, **490**, 239–253.
 11. Helmig, D., Daly, R. W., Milford, J. and Guenther, A., Seasonal trends of biogenic terpene emissions. *Chemosphere*, 2013, **93**(1), 35–46.
 12. Penuelas, J. and Staudt, M., BVOCs and global change. *Trends in Plant Sci.*, 2010, **15**(3), 133–144.
 13. Csiky, O. and Seufert, G., Terpenoid emissions of Mediterranean oaks and their relation to taxonomy. *Ecol. Appl.*, 1999, **9**(4), 1138–1146.
 14. Dudareva, N., Pichersky, E., Negre, F. and Gershenzon, J., Biochemistry of plant volatiles. *Plant physiol.*, 2004, **135**(4), 1893–1902.
 15. Karl, T., Guenther, A., Turnipseed, A., Tyndall, G., Artaxo, P. and Martin, S., Rapid formation of isoprene photo-oxidation products observed in Amazonia. *Atmos. Chem. Phys.*, 2009, **9**(20), 7753–7767.
 16. Llusia, J., Penuelas, J. and Gimeno, B. S., 2002. Seasonal and species-specific response of VOC emissions by Mediterranean woody plant to elevated ozone concentrations. *Atmos. Environ.*, **36**(24), 3931–3938.
 17. Malik, T. G., Gajbhiye, T. and Pandey, S. K., Plant specific emission pattern of biogenic volatile organic compounds (BVOCs) from common plant species of Central India. *Environ. Monit. Assess.*, 2018a, **190**(11), 1–11.
 18. Penuelas, J. and Llusia, J., BVOCs: plant defense against climate warming? *Trends in Plant Sci.*, 2003, **8**, 105–109.
 19. Guenther, A., Karl, T., Harley, P., Wiedinmyer, C., Palmer, P. I. and Geron, C., Estimates of global terrestrial isoprene emissions using MEGAN (Model of Emissions of Gases and Aerosols from Nature). *Atmos. Chem. Phys.*, 2006, **6**, 3181–3210, <http://www.atmos-chem-phys.net/6/3181/2006/>
 20. Karl, T., *et al.*, The tropical forest and fire emissions experiment: Emission, chemistry, and transport of biogenic volatile organic compounds in the lower atmosphere over Amazonia, *J. Geophys. Res.*, 2007, **112**, D18302, doi:10.1029/2007JD008539.
 21. Stibig, H. J., Achard, F., Carboni, S., Rasi, R. and Miettinen, J., Change in tropical forest cover of Southeast Asia from 1990 to 2010. *Biogeosciences*, 2014, **11**, 247–258.

22. Malik, T. G., Gajbhiye, T. and Pandey, S. K., Seasonality in emission patterns of isoprene from two most dominant tree species of Central India: Implications on terrestrial carbon emission and climate change. *Proc. Int. Acad. Ecol. Environ. Sci.*, 2018b, **8**(4), 204-212.
23. Malik, T. G., Gajbhiye, T. and Pandey, S. K., Some insights into composition and monoterpene emission rates from selected dominant tropical tree species of Central India: Plant-specific seasonal variations. *Ecol. Res.*, 2019, **34**(6), 821-834. doi:10.1111/1440-1703.12058.
24. Singh, R., Singh, A. P., Kumar, A., Singh, M. P. and Varshney, C. K., Emission of isoprene from common Indian plant species and its implications for regional air quality. *Environ. Monit. Assess.*, 2008, **144**, 43–51.
25. Singh, A. P., Singh, R., Mina, U., Singh, M. P. and Varshney, C. K., Emissions of monoterpene from tropical Indian plant species and assessment of VOC emission from the forest of Haryana state. *Atmos. Pollut. Res.*, 2011, **2**(1), 72-79.
26. Varshney, C. K. and Singh, A. P., Isoprene emission from Indian trees. *J. Geophys. Res. Atmos.*, 2003, **108** (D24), 4803. doi:10.1029/ 2003JD003866.
27. FSI: State of Forest Report, Forest Survey of India, Ministry of Environment and Forest, Government of India, 2021.
28. ISFR: India State of Forest Report, Forest Survey of India Ministry of Environment, Forest and Climate Change Government of India. 2017, Vol. I.
29. Roy, P. S., *et al.*, New vegetation type map of India prepared using satellite remote sensing: Comparison with global vegetation maps and utilities. *Int. J. Appl. Earth Obs. Geoinf.*, 2015, **39**, 142-159.
30. <https://bhuvan.nrsc.gov.in>
31. Padhy, P. K. and Varshney, C. K., Emission of volatile organic compounds (VOC) from tropical plant species in India. *Chemosphere*, 2005a, **59**(11), 1643-1653.
32. Padhy, P. K. and Varshney, C. K., Isoprene emission from tropical tree species. *Environ. Pollut.*, 2005b, **135**(1), 101-109.
33. Singh, A. P. and Varshney, C. K., Isoprene emission from the forest of Haryana state. *Environ. Monit. Assess.*, 2006, **122**(1), 145-151.
34. Singh, A. P., Varshney, C. K., Padhy, U. and Singh, U. K., Seasonal variations in isoprene emission from Indian tropical deciduous tree species. *Environ. Monit. Assess.*, 2007, **131**(1-3), 231-235.

35. Singh, R., Singh, M. P. and Singh, A. P., Ozone forming potential of tropical plant species of the Vidarbha region of Maharashtra state of India. *Urban For. Urban Green.*, 2014, **13**(4), 814-820.
36. Helmig, D., Ortega, J., Guenther, A., Herrick, J. D. and Geron, C., Sesquiterpene emissions from loblolly pine and their potential contribution to biogenic aerosol formation in the Southeastern US. *Atmos. Environ.*, 2006, **40**, 4150–4157.
37. Kim, J. C., Factors controlling natural VOC emissions in a southeastern US pine forest. *Atmos. Environ.*, 2001. **35**(19), 3279-3292.
38. Schuh, G., Heiden, A. C., Hoffmann, T., Kahl, J., Rockel, P., Rudolph, J. and Wildt, J., Emissions of volatile organic compounds from sunflower and beech: dependence on temperature and light intensity. *J. Atmos. Chem.*, 1997, **27**(3), 291-318.
39. Lin, C. Y., Chang, T. C., Chen, Y. H., Chen, Y. J., Cheng, S. S. and Chang, S. T., Monitoring the dynamic emission of biogenic volatile organic compounds from *Cryptomeria japonica* by enclosure measurement. *Atmos. Environ.*, 2015, **122**, 163-170.
40. Niinemets, U. *et al.*, Estimations of isoprenoid emission capacity from enclosure studies: data processing, quality and standardized measurement protocols. *Biogeosciences*, 2011, **8**, 2209–2246.
41. Ortega, J. and Helmig, D., Approaches for quantifying reactive and low-volatility biogenic organic compound emissions by vegetation enclosure techniques–Part A' *Chemosphere*, 2008, **72**, 343–364.
42. Tsui, J. K. Y., Guenther, A., Yip, W. K. and Chen, F., A biogenic volatile organic compound emission inventory for Hong Kong. *Atmos. Environ.*, 2009, **43**, 6442-6448.
43. Préndez, M., Carvajal, V., Corada, K., Morales, J., Alarcón, F. and Peralta, H., Biogenic volatile organic compounds from the urban forest of the Metropolitan Region, Chile. *Environ. Pollut.*, 2013, **183**, 143-150.
44. Komenda, M. and Koppmann, R., Monoterpene emissions from Scots pine (*Pinus sylvestris*): field studies of emission rate variabilities. *J. Geophys. Res. Atmos.*, 2002, **107**(D13), 4161, 10.1029/2001JD000691.
45. Lüpke, M., Steinbrecher, R., Leuchner, M. and Menzel, A., The tree drought emission monitor (tree demon), an innovative system for assessing biogenic volatile organic compounds emission from plants. *Plant Methods*, 2017, **13**(1), 1–17. <https://doi.org/10.1186/s13007-017-0166-6>.

46. Chen, J., Bi, H., Yu, X., Fu, Y. and Liao, W., Influence of physiological and environmental factors on the diurnal variation in emissions of biogenic volatile compounds from *Pinus tabuliformis*. *J. Environ. Sci. (China)*, 2019, **81**, 102–118. <https://doi.org/10.1016/j.jes.2019.01.020>.
47. Huang, X., *et al.*, Biogenic volatile organic compound emissions from *Pinus massoniana* and *Schima superba* seedlings: Their responses to foliar and soil application of nitrogen. *Sci. Total Environ.*, 2020, **705**, 135761, <https://doi.org/10.1016/j.scitotenv.2019.135761>.
48. Mozaffar, A., *et al.*, Methanol emissions from maize: Ontogenetic dependence to varying light conditions and guttation as an additional factor constraining the flux. *Atmos. Environ.*, 2017, **152**, 405-417.
49. Bracho-Nunez, A., Welter, S., Staudt, M. and Kesselmeier, J., Plant-specific volatile organic compound emission rates from young and mature leaves of Mediterranean vegetation. *J. Geophys. Res. Atmos.*, 2011, **116**(D16304), doi: [10.1029/2010JD015521](https://doi.org/10.1029/2010JD015521).
50. Llusia, J., Penuelas, J., Sardans, J., Owen, S. M. and Niinemets, U., Measurement of volatile terpene emissions in 70 dominant vascular plant species in Hawaii: aliens emit more than natives. *Glob. Ecol. Biogeogr.*, 2010, **19**(6), 863-874.
51. Llusia, J., Sardans, J., Niinemets, Ü., Owen, S. M. and Peñuelas, J., A screening study of leaf terpene emissions of 43 rainforest species in Danum Valley Conservation Area (Borneo) and their relationships with chemical and morphological leaf traits. *Plant Biosyst.*, 2013, **148**(2), 307-317. doi:10.1080/11263504.2013.770803.
52. Sardans, J., Llusia, J., Owen, S. M., Niinemets, U. and Penuelas, J., Screening study of leaf terpene concentration of 75 Borneo rainforest plant species: relationships with leaf elemental concentrations and morphology. *Rec. Nat. Prod.*, 2015, **9**(1), 19-40.
53. Aksoyoglu, S., *et al.*, Aerosol modelling in Europe with a focus on Switzerland during summer and winter episodes. *Atmos. Chem. Phys.*, 2011, **11**(14), 7355-7373.
54. Steinbrecher, R., *et al.*, Intra-and inter-annual variability of VOC emissions from natural and semi-natural vegetation in Europe and neighbouring countries. *Atmos. Environ.*, 2009, **43**(7), 1380-1391.
55. Srivathsa, A., Karanth, K. U., Kumar, N., and Oli, M. K., Insights from distribution dynamics inform strategies to conserve a dhole *Cuon alpinus* metapopulation in India. *Scientific Reports*, 2019, **9**(1), 1-12.

56. FSI, State of Forest Report, Forest Survey of India, Ministry of Environment and Forest, Government of India, 1987.
57. Rao, R. R., Floristic diversity in Western Ghats: Documentation, Conservation and Bioprospection—A Priority Agenda for Action, 2013.
58. FDGG, Forest Department, Government of Goa, Ministry of Environment, Forest and Climate Change, 2022.
59. Yaman, B., *et al.*, Biogenic volatile organic compound (BVOC) emissions from various endemic tree species in Turkey. *Aerosol Air Qual. Res.*, 2015, **15**(1), 341-356.
60. Sahu, L. K., Tripathi, N., Gupta, M., Singh, V., Yadav, R. and Patel, K., Impact of COVID-19 pandemic lockdown in ambient concentrations of aromatic volatile organic compounds in a metropolitan city of western India. *J. Geophys. Res. Atmos.*, 2022, **127**(6), <https://doi.org/10.1029/2022JD036628>.
61. Hakola, H., Rinne, J., and Laurila, T., The hydrocarbon emission rates of tea-leaved willow (*Salix phylicifolia*), silver birch (*Betula pendula*) and European aspen (*Populus tremula*). *Atmos. Environ.*, 1998, **32**, 1825–1833.
62. Fiorani, F., Bogemann, G. M., Visser, E. J. W., Lambers, H., and Voesenekm, L. A. C. J., Ethylene emission and responsiveness to applied ethylene vary among Poa species that inherently differ in leaf elongation rates. *Plant Physiol.*, 2002, **129**, 1382–1390. doi: 10.1104/pp.001198.
63. Reid, M. S., Ethylene in plant growth, development, and senescence. In *Plant Hormones*, (ed. P. J. Davis), Dordrecht, Springer, 1995, 486-508, doi:10.1007/978-94-011-0473-9_23.
64. Lin, Z. F., Zhong, S. L. and Grierson, D., Recent advances in ethylene research. *J. Exp. Bot.*, 2009, **60**, 3311–3336.
65. Qi, X., Liu, C., Song, L., and Li, M. PaMADS7, a MADS-box transcription factor, regulates sweet cherry fruit ripening and softening. *Plant Sci.*, 2020, **301**, 110634. <https://doi.org/10.1016/j.plantsci.2020.110634>.
66. Okumura, M., Kosugi, Y. and Tani, A., Biogenic volatile organic compound emissions from bamboo species in Japan. *J. Agric. Meteorol.*, 2018, **74**(1), 40-44.
67. Funk, J. L., Physiological and environmental controls over isoprene emission. (2004). UMI Number: 3131286, State University of New York at Stony Brook.

492 Table1. Concentrations (mean \pm standard deviation) of different BVOCs and values of
 493 environmental parameters measured during the chamber experiments from seven different
 494 plant species.

Plant species	Ethene (ppbv)	Propene (ppbv)	Isoprene (ppbv)	α -pinene (ppbv)	β -pinene (ppbv)	Sampling date and time	T (°C)	RH (%)	Solar radiation (W/m ²)
<i>Saraca asoca</i>	0.73 \pm 0.19	0.41 \pm 0.12	0.74 \pm 0.09	0.03 \pm 0.008	0.03 \pm 0.006	12:55 (16 October 2022)	29.22 \pm 0.16	69	591.25 \pm 35.38
<i>Leucaena leucocephala</i>	0.53 \pm 0.11	0.45 \pm 0.12	0.34 \pm 0.11	0.02 \pm 0.007	0.02 \pm 0.004	12:05 (14 October 2022)	29.05 \pm 0.38	62	620.08 \pm 12.98
<i>Psidium guajava</i>	0.39 \pm 0.14	0.44 \pm 0.19	6.64 \pm 1.82	0.01 \pm 0.001	0.01 \pm 0.007	13:20(14 October 2022)	28.23 \pm 0.15	70	459.83 \pm 77.11
<i>Gliricidia sepium</i>	NA	NA	NA	0.02 \pm 0.01	0.03 \pm 0.001	15:10(13 October 2022)	27.96 \pm 0.19	67	216.90 \pm 112.82
<i>Manilkara zapota</i>	0.9	0.49	0.31	0.01	0.01	11:45 (13 October 2022)	28.39 \pm 0.52	76	625.34 \pm 15.42
<i>Bambusa vulgaris</i>	0.88	0.83	0.65	0.03	0.04	11:30 (16 October 2022)	29.66 \pm 0.30	63	707.20 \pm 12.38
<i>Tectona grandis</i>	0.92	0.60	6.84	0.01	0.01	12:50 (13 October 2022)	28.33 \pm 0.17	76	493.69 \pm 60.11

495

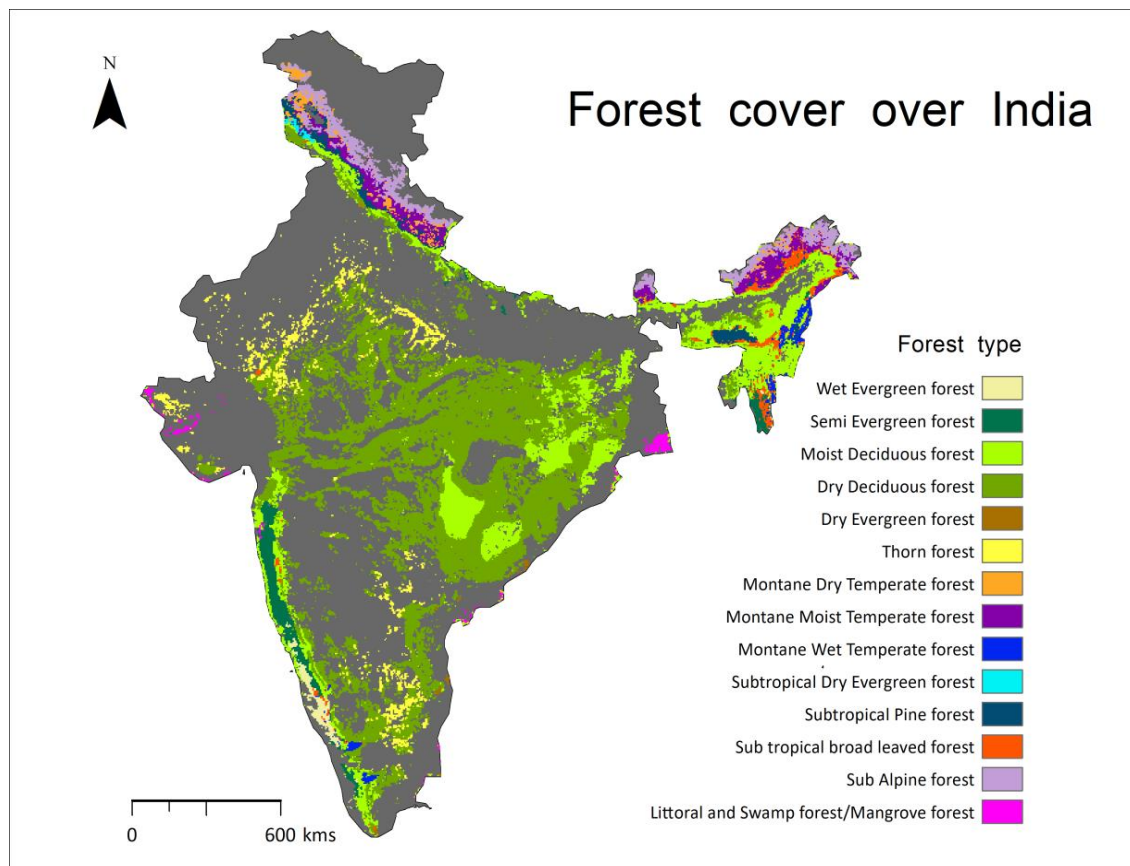


Figure 1. Different forest types and land use/land cover map of India using Forest type 5km grid data, National Remote Sensing Centre, ISRO, Government of India, Hyderabad, India, through the Bhuvan Geo Portal (<https://bhuvan.nrsc.gov.in>)

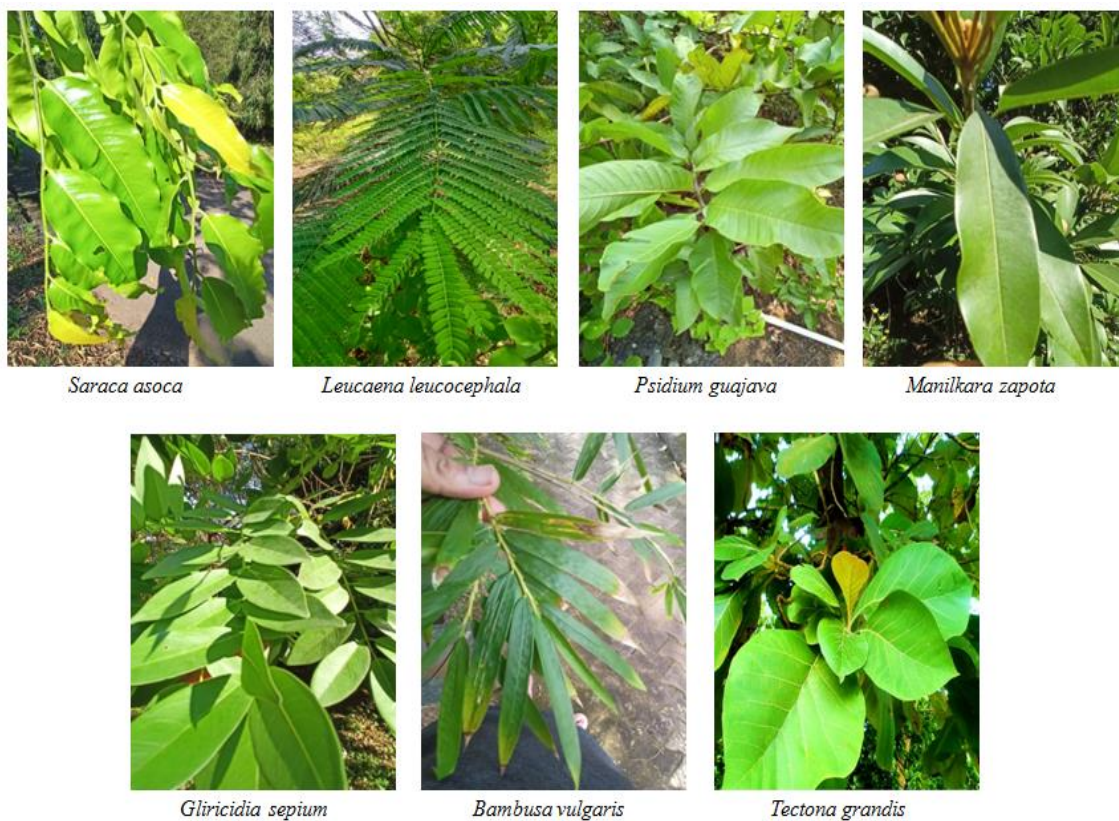


Figure 2. Pictures showing the leaves of seven different plant species selected for the chamber experiments which are dominant species in the Western Ghats of India.

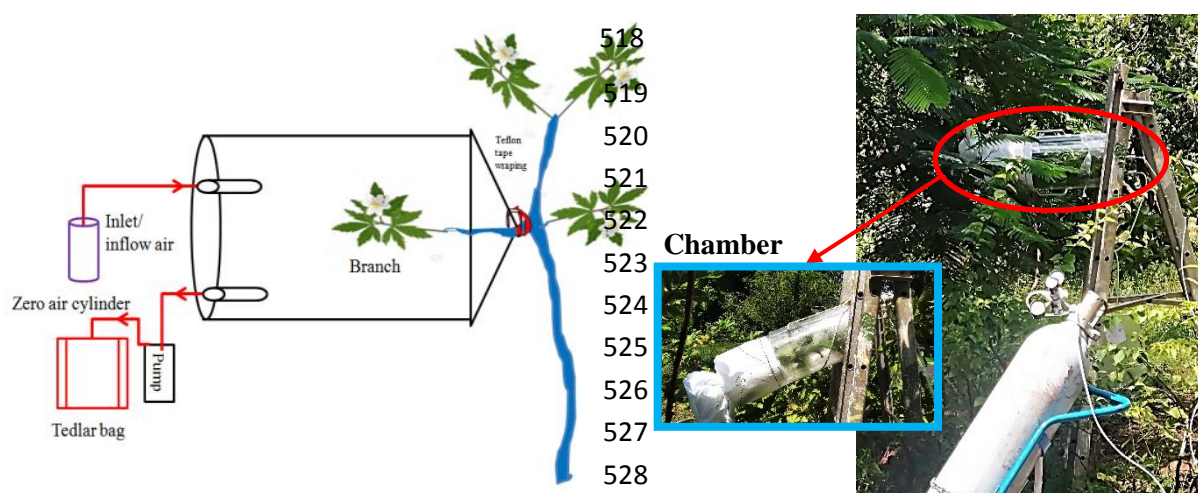


Figure 3. Schematic diagram (left) and field deployment (right) of the dynamic chamber system for the sampling of BVOCs emitted from different plant species.

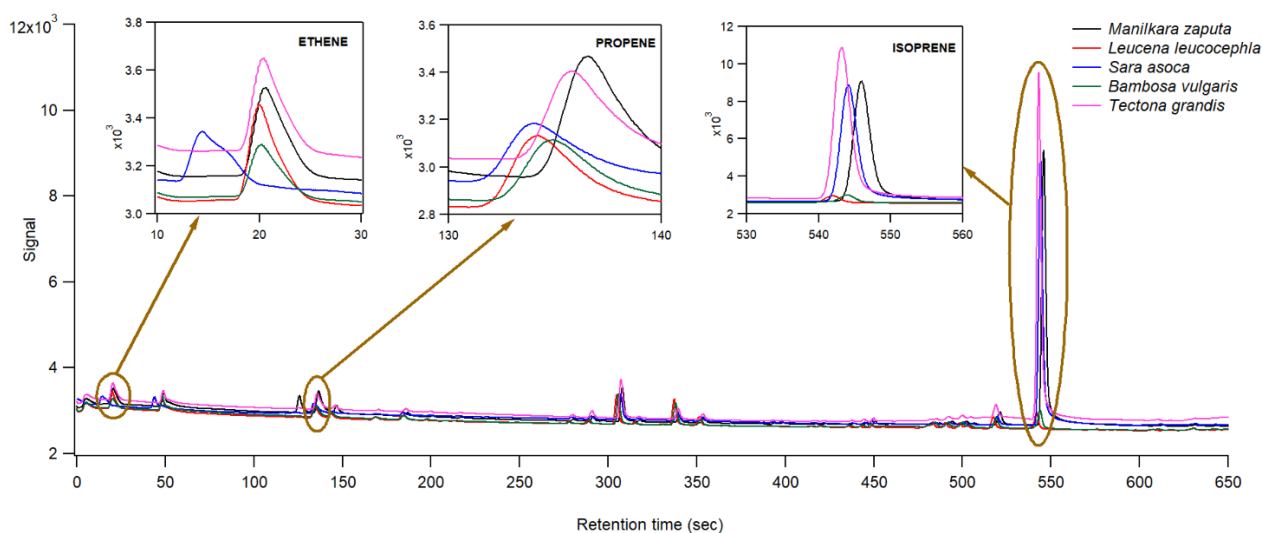


Figure 4. Typical chromatograms obtained from the analysis of samples collected from the chamber experiments on selected plant species.

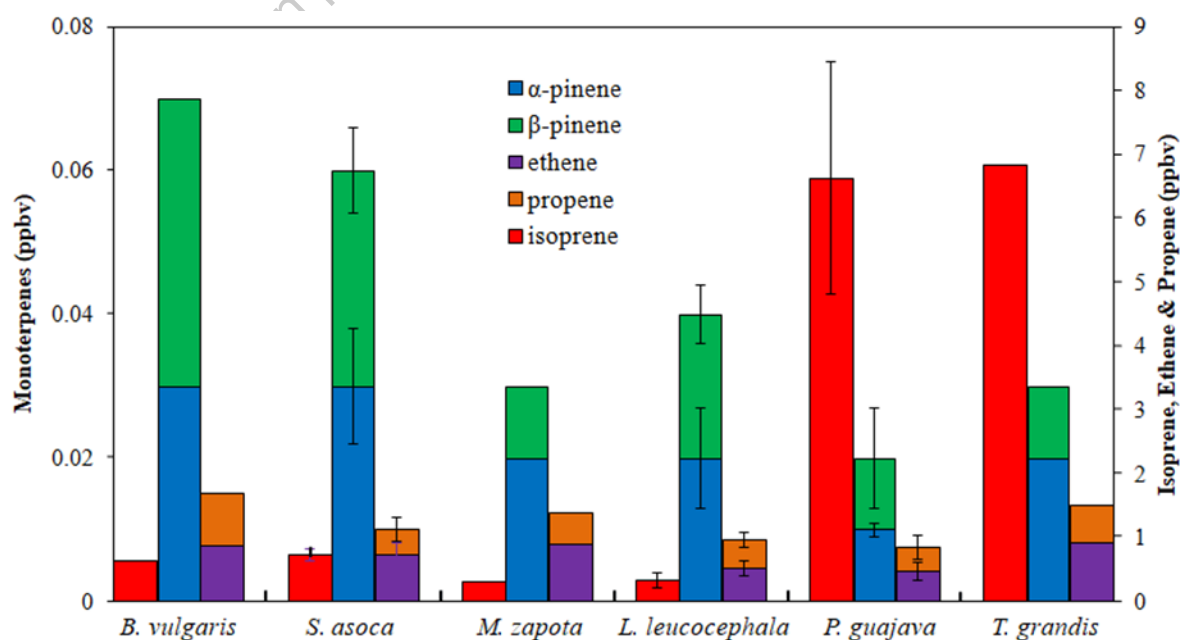


Figure 5. The concentrations (mean) of alkenes (ethene and propene), isoprene and monoterpenes (α -pinene & β -pinene) in the samples collected from plant species during the chamber experiments, the error bars represent the standard deviation.